

REVIEW

Open Access



Recent advances in modified poly (lactic acid) as tissue engineering materials

Samanta Castañeda-Rodríguez^{1,2}, Maykel González-Torres^{1,2*}, Rosa María Ribas-Aparicio²,
María Luisa Del Prado-Audelo³, Gerardo Leyva-Gómez⁴, Eda Sönmez Güler⁵ and Javad Sharifi-Rad^{6*}

Abstract

As an emerging science, tissue engineering and regenerative medicine focus on developing materials to replace, restore or improve organs or tissues and enhancing the cellular capacity to proliferate, migrate and differentiate into different cell types and specific tissues. Renewable resources have been used to develop new materials, resulting in attempts to produce various environmentally friendly biomaterials. Poly (lactic acid) (PLA) is a biopolymer known to be biodegradable and it is produced from the fermentation of carbohydrates. PLA can be combined with other polymers to produce new biomaterials with suitable physicochemical properties for tissue engineering applications. Here, the advances in modified PLA as tissue engineering materials are discussed in light of its drawbacks, such as biological inertness, low cell adhesion, and low degradation rate, and the efforts conducted to address these challenges toward the design of new enhanced alternative biomaterials.

Keywords Biocomposite, Biotechnology, Fabrication, Nanotechnology, Poly (lactic acid), Tissue Engineering

Introduction

In the past few years, the regenerative medicine area has improved several achievements in looking forward to materials suitable for tissue engineering. A paramount concern of humanity is the implementation of techniques that favor the environment; such concerns

have led us to develop materials with a lower ecological footprint. In a tangible field such as the environment in which we develop, there are factors to consider in reducing waste that, in turn, affect our ecological footprint on the planet. Besides, using toxic materials and biological waste for research is a latent problem [1]. To reduce the environmental impact and with the urgency of developing new materials, it has been sought to use polymers from natural and renewable sources extracted from bacteria, plants, or other organisms to implement innovative technologies such as tissue engineering with exponential growth in recent years.

Renewable resources have been sought to develop new materials, resulting in attempts to produce a wide variety of biomaterials that are friendly to the environment and low cost. However, the challenge in producing these new materials is that they must be biocompatible with organisms. In the field of tissue engineering, the biomaterials that are developed must be biocompatible and have a specific mechanical resistance to function as a support

*Correspondence:

Maykel González-Torres
maykel.gonzalez@conacyt.mx
Javad Sharifi-Rad
javad.sharifirad@gmail.com

¹Conacyt & Laboratorio de Biotecnología, Instituto Nacional de Rehabilitación, Ciudad de México, México

²Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional (IPN), Ciudad de México, México

³Escuela de Ingeniería y Ciencias, Tecnológico de Monterrey, Campus Ciudad de México, México

⁴Departamento de Farmacia, Facultad de Química, Universidad Nacional Autónoma de México, Ciudad de México, México

⁵Faculty of Pharmacy, Department of Pharmacognosy, Sivas Cumhuriyet University, Sivas, Turkey

⁶Facultad de Medicina, Universidad del Azuay, Cuenca, Ecuador



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

during the repair of damaged tissue. Using natural materials such as biopolymers can offer benefits for various applications, enhancing their properties, which are convenient for studying and treating diseases [2]. Poly (lactic acid) (PLA) is a biopolymer known to be eco-friendly, which has the characteristic of being a biodegradable polymer produced from the fermentation of carbohydrates. This characteristic allows it to be produced on a large scale and with reduced costs. Also, its production has low emissions of greenhouse gases [3]. This characteristic of biodegradability has been of great interest to tissue engineering due to its application for developing scaffolds and nanomaterials.

Moreover, the polymer industry kept growing until we were aware of these years, and PLA has been modified using various synthetic, semi-synthetic, or natural polymers to enhance its properties and thus design copolymers with new applications beneficial for biomedicine. Because of these advances, new applications have been updated, and new challenges for tissue engineering have been developed.

Recently, producing PLA and PLA-based derivatives for medical applications has received growing attention [4]. The use of emulsion, wet, blend, and coaxial electrospinning from PLA-based structures and their biomedical applications were reviewed over the last five years for the period up to the present day [5]. Similarly, a concise review focused on current kidney tissue engineering applications of PLA electrospun scaffolds [6]. Also, the review of the effect of PLA processing conditions on the physicochemical and biological material properties [7], the use of PLA-based microparticles for drug delivery [8], the use of PLA composites and blends for cutting-edge biotechnologies [9], PLA 3D printing [10], PLA membranes synthesis [11], and bioactive coatings of PLA for bone tissue engineering were highlighted and recently conducted [12]. The above reviews demonstrate that the topic of PLA derivatives designed to be used in biomedicine is fascinating, of much interest, and practical usefulness. However, the reviews carried out are particular for each topic, and to our knowledge, no review has collected the most recent and interesting works on the different PLA derivatives as well as the most suitable synthesis techniques for the use of this versatile biomaterial in tissue engineering, which is the main novelty and relevance of this work. Therefore, studying, highlighting and emphasizing the PLA intriguing history, the biosynthesis, the search of the different types of modifications and derivatives, as well as the recent advances of the most promising strategies for PLA use in tissue engineering is presented here.

Inside the history of polymers: poly (lactic acid)

Lactic acid (LA) was discovered in 1780 by the Swedish chemist Carl Scheele from sour milk, and years later, Jöns Berzelius, in 1808, discovered the L-lactic acid, better known as L-Lactate, a molecule produced in muscles [13, 14]. According to Dorgan et al. [15], in 1832, Wallace Carothers developed the PLA when they tried to polymerize and depolymerize oligomeric lactides by polycondensation [16]. After that, in 1954, the PLA synthesis was improved to produce a high molecular weight, but it was costly. In 1966, Kulkarni et al. [17] established that PLA is a nontoxic, non-tissue-reactive, and slowly degrading compound that is possibly entirely metabolized through the respiratory system. Such discoveries have been the very beginning of the biomedical applications of PLA. As a result of the novel applications, the production of PLA increased to what we know today [Fig. 1].

Lactic acid and poly (lactic acid)

Poly (lactic acid) is an organic polymer derived from lactic acid, a chemical compound within organisms. As a result, lactic acid is one of the most important molecules in our bodies because it is a precursor in several metabolic pathways and is produced by animals, plants, and microorganisms. Besides, LA can be a good component of synthesizing other compounds because of the functional groups with which the lactic acid counts. The chemical structure of LA can be numbered in the hydroxyl and carbonyl groups. The LA, in its ionic form, is called lactate. Also, in the IUPAC nomenclature, the complete name is 2-hydroxypropanoic acid, a carboxylic acid with a hydroxy group in the α carbon, and the condensate formula is $(\text{CH}_3\text{-CHOHCOOH})$ [18] [Fig. 2a]. Besides, lactic acid is a molecule with optical activity, which counts with a racemic mixture. The LA has three enantiomeric forms, which are L (+), D (-), and LD (+/-) [Fig. 2b and c]. In addition, the pure mixture of L-lactic acid and D-lactic acid has a high commercial value in the industry, and in fact, L-lactic acid is the chemical structure that is the monomer of poly (lactic acid) [19].

Metabolic pathways and biosynthesis of the lactic acid

Lactic acid, from being a small organic molecule present in various organisms and participating in many biochemical processes, has the quality of not only being produced in the human body but also in microorganisms. In addition, it has given the possibility that lactic acid can be produced through biotechnological processes, which has allowed the advancement of the incorporation of biotechnologically modified strains or microorganisms for the development of new products [20–25].

Several microorganisms have been able to produce lactic acid and organic lactates, such as fungi [26, 27],

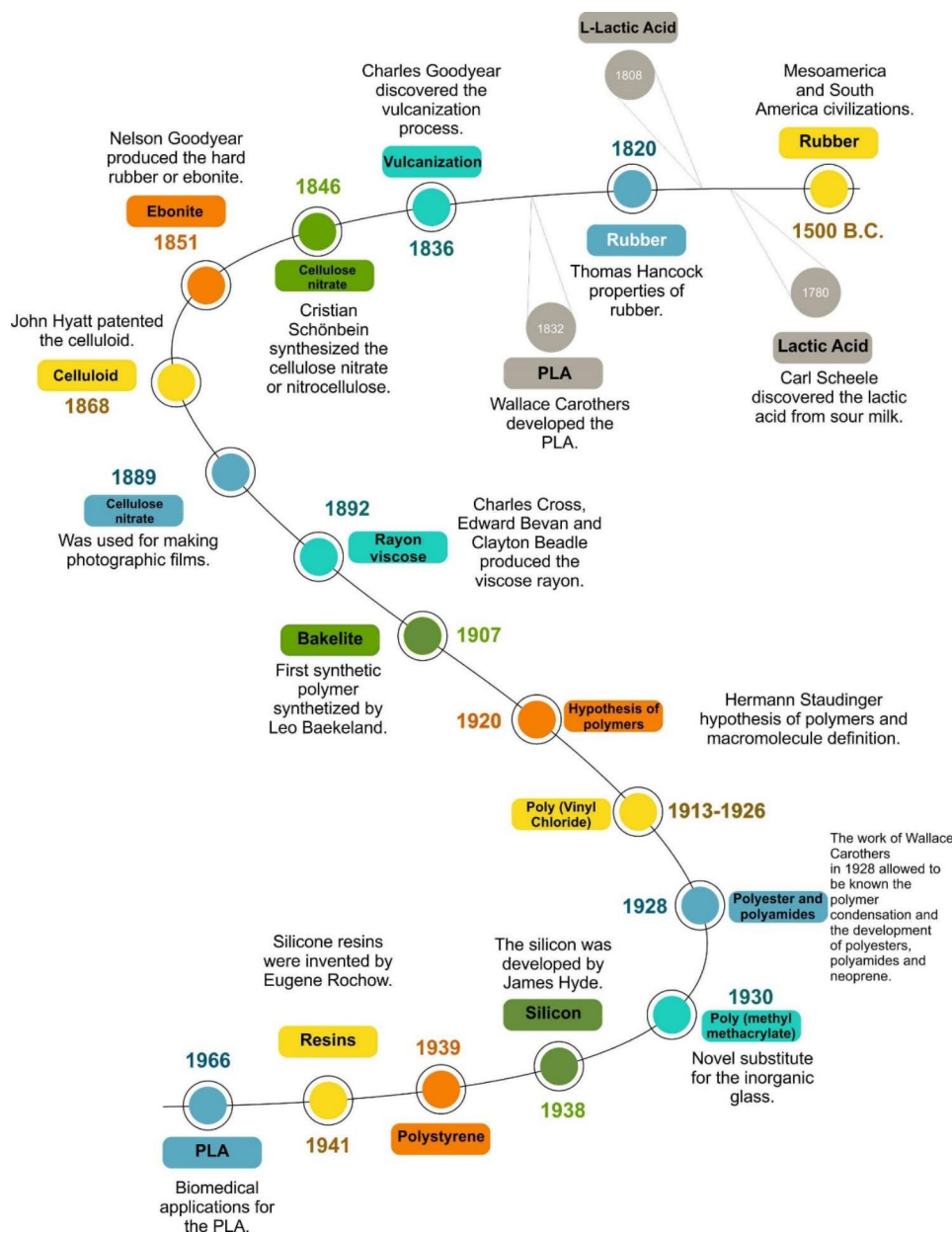


Fig. 1 Inside the history of polymers

cereal [28], yeast [29–31], cyanobacteria [32–35], and even algae [36], despite the great variety of microorganisms that produce LA, bacteria have been the most commonly used for the biotechnological production of lactic acid due to the ease and versatility of the handling and processing of bacteria. One of the most important genera of bacteria for producing lactic acid is the genera *Lactobacillus*. On the other hand, other bacteria produce LA, such as the *Bifidobacterium* genera, *Bacillus Sporolactobacillus*. Although these bacteria are not found within the group of lactic acid-producing genera, they include *Enterococcus faecium*, *Lactococcus lactis*, *Pediococcus acidilactici*, and *Streptococcus thermophilus* produce

LA [37, 38]. Table 1 summarizes the most common lactic acid-producing bacteria and some of their characteristics.

According to Carr *et al.* [37], lactic acid bacteria can be classified into Homofermenters and Heterofermenters based on the type of production the bacteria can make. The homofermenters can produce LA by taking glucose as a product and transforming it by oxidation and fermentation. Moreover, the heterofermenters can produce other products instead the lactic acid; such products can be acetic acid, CO₂, and ethanol produced by the fermentation of the glucose [Fig. 3]. It is worth mentioning that lactic acid bacteria are a broad group of gram-positive bacteria and are obligate fermentative

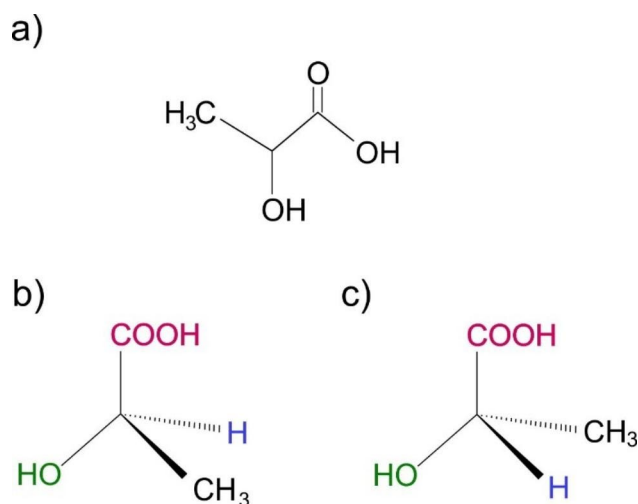


Fig. 2 Structure of lactic acid. (a) The chemical structure of lactic acid (C₃H₆O₃), the IUPAC name is 2 – Hydroxypropanoic acid. (b) Isomeric structure of lactic acid, (R) or L (+) Lactic acid. (c) Isomeric structure of lactic acid, (S) or D (-) Lactic acid

[39]. The homofermentative bacteria can reduce hexose carbohydrates by the glycolysis pathway; the glycolysis breakdown the molecules of glucose or hexoses to turn them into pyruvate, and the primary production of lactic acid is generated by the enzymes lactate dehydrogenases (LDH) (EC 1.1.1.27).

Also, the LDH enzyme family can exist in an L or D stereospecific form [43]. In addition, those enzymes are oxidoreductases, and the LDHs can be classified into two groups, the NAD-dependent LDH, and the NAD-independent LDH. LDH NAD-dependent enzymes catalyze the reaction. In other words, they depend on NADH's oxidation to NAD⁺. The L-LDH NAD-dependent catalyzes a redox reaction in which the pyruvate is reduced into L-lactate or L-lactic acid; those enzymes can generate the reaction reversibly or irreversibly. In this, the final product of glycolysis is lactate. According to Garvie [44],

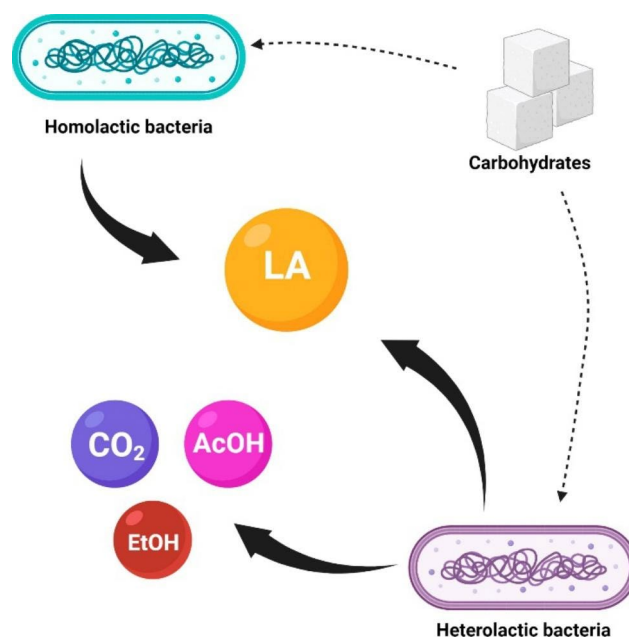


Fig. 3 Products of fermentation of heterolactic and homolactic bacteria

the LDH enzymes can differ between species existing in different stereoisomer forms for the L or D forms, dependent or non-dependent forms.

In contrast with the homolactic fermentative bacteria, under anaerobic conditions, the heterolactic fermentative bacteria use the phosphogluconate path, commonly known as the phosphoketolase pathway, to earn LA, acetic acid, ethanol, and CO₂ [45–47]. The phosphogluconate pathway transforms the glucose or hexoses into pentoses through the enzyme phosphoketolase (EC 4.1.2.22) [48]. The enzyme catalyzes a reaction producing glyceraldehyde 3-phosphate (G3P) and acetyl phosphate [Fig. 4]. If a pentose enters, the path does not yield CO₂ [49, 50]. The phosphoketolases (Pkts) catalyze an irreversible reaction and play together with other enzymes, like acetate kinase and phosphotransacetylases, to

Table 1 Summary of the most common lactic acid-producing bacteria and some of their characteristics

Genus	Shape	Metabolism	Microorganism	Growing conditions	Culture media	Refs.
Lactobacillus	Bacilli/pairs/chains	Homofermentative Heterofermentative	L. acidophilus	T° opt.: 30–40 °C (2–53 °C) pH opt.: 5.5–6.2, tolerant < 4	Requires individually various complex nutritional requirements for peptides, amino acids, nucleotides, vitamins, and fermentable carbohydrates	[20, 22, 40, 41]
			L. delbrueckii			
			L. brevis			
			L. fermentis			
Lactococcus	Cocci/chains	Homofermentative	L. lactis spp. lactis	T° opt: 10 °C < 45 °C.	It may be selectively isolated on Elliker's lactic agar, Arginine Tetrazolium Agar, or Alsan Medium. They usually grow in media containing 4% (w/v) NaCl	
			L. lactis spp. cremoris			
Pediococcus	Cocci/tetrad	Homofermentative	P. acidilacti	Reduced atmospheric conditions	Pediococci grows on MRS media, and growth may be enhanced, as with the Leuconostocs.	[20, 22, 40, 42]
			P. cellicola			
			P. clausenii			
Leuconostoc	Pairs/chains	Heterofermentative	L. mesenteroides	Alkaline environment, pH opt: ≥ 4.5.	Although MRS agar is suitable for Leuconostocs, Yeast Glucose Phosphate Peptone Broth is recommended.	
			L. cremoris			
			L. oenos			

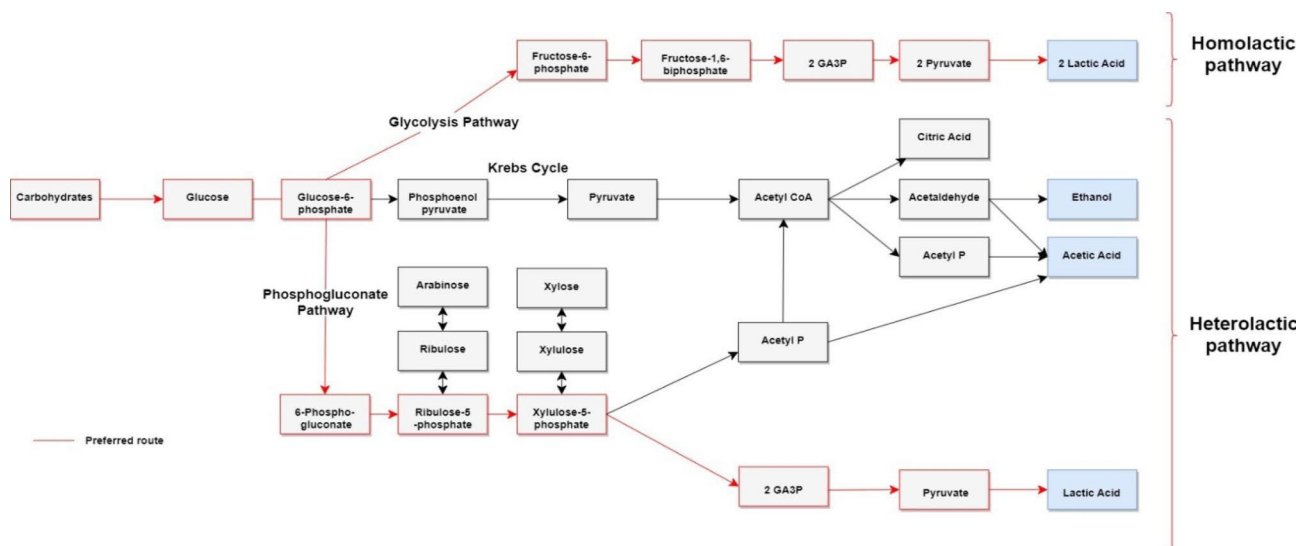


Fig. 4 Summarized homolactic and heterolactic pathways of lactic acid bacteria

produce acetate, ATP, and acetyl Co-A [51]. Also, G3P is oxidated in pyruvate and lactic acid. On the other hand, at the end of the reaction, the acetyl phosphate turns into ethanol [22, 52, 53]. In humans, as in homofermentative bacteria, the lactate is produced by glycolysis using glucose to transform it into pyruvate and, by the action of the LDH, is converted into lactate by a reversible reaction with NADH as a coenzyme. In the human body, lactate is a waste product of anaerobic metabolisms and is used in gluconeogenesis to develop energy by oxidation in metabolic pathways like the Krebs cycle and Cory cycle; as a result, the skeletal muscle is the first consumer of lactate in the organism [54].

Also, the accumulation of lactate in the organisms, to be more specific, the accumulation of lactate in the blood, can bring consequences to the health and the generation of lactic acidosis [55–57].

Polymerization of poly (lactic acid)

Two different methods can prepare the polymerization of PLA. The first method is polycondensation, and according to Garlotta [58], the polymerization of lactic acid results in a low molecular weight polymer, brittle, glassy polymer, and is unusable unless mixed with other substances to increase its molecular weight.

In addition, the molecular weight of this polymer is low due to certain factors such as viscosity, impurities, water in the molecule, and the low concentration of reactive end-groups. In the polycondensation method, as its name indicates, water is removed from the solution by condensation using the solvent under elevated temperature and high vacuum conditions; a polycondensation reaction in regular terms is conducted in bulk via distillation by condensation of water. Likewise, the transesterification process is accelerated if a reaction catalyst is added. This

lactic acid polymerization method allows the synthesis of PLA oligomers. Although the average molecular weight of the polymer is low compared to other methods mentioned, it allows the addition of other compounds to increase the molecular weight of the polymer [59]. The second method consists of generating a new molecule whose function is to be a cyclical intermediary; this intermediary is called lactide. The polymerization method is by ring-open polymerization (ROP) where under specific conditions, such as heat and without the need for a solvent, a high molecular weight polymer is obtained; this method allows the polymerization process to be conducted, which favors us in obtaining a pure polymer, guaranteeing a higher yield of the reaction. According to Metha et al. [60], a catalyst frequently utilized to conduct this reaction is stannous octoate from zinc metal. In addition, the choice of catalyst, initiator, and co-initiator for this ROP reaction affects the properties of the PLA polymer. A disadvantage of this type of polymerization is that conducting this process is unfavorable due to the high costs of polymerization. However, the development of innovative technologies such as membrane design, ultrafiltration, chromatography, and electro dialysis, among others, has allowed purification costs to decrease the PLA and therefore improve the processes, making it possible to obtain more efficient products. Figure 5 illustrates the types of polymerizations of polylactic acid [18] [Fig. 5].

Characteristics and properties of PLA

PLA has a wide range of characteristics and properties. Hagen [25] describes that PLA is a transparent glass that is opaque when crystallized. In addition, Table 2 summarizes some characteristics and properties of PLA.

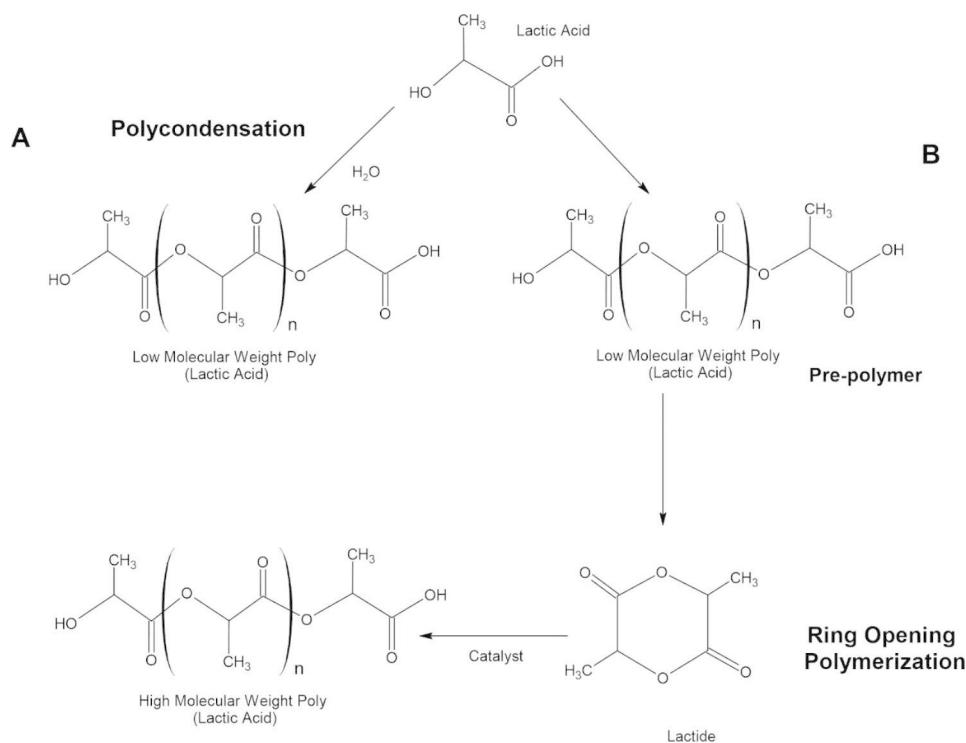


Fig. 5 Synthesis of poly (lactic acid). (A) The polymerization of lactic acid by polycondensation results in a low molecular weight polymer, water is removed from the solution under elevated temperature and high vacuum conditions. (B) The generation of Lactide a cyclical intermediary, allows the ring-open polymerization (ROP) where under specific conditions (heat and accurate solvent) a high molecular weight polymer is obtained, with ROP a pure polymer and a higher yield of the reaction is obtained

Table 2 Properties of poly (lactic acid)

Properties	Value	Refs.
Melting temperature	$T_m = 230 < 240$ °C	[61, 62]
Equilibrium T_m	$T_m = 165\text{--}279$ °C	[61–63]
Glass transition temperature (T_g)	$T_g = 65\text{--}72$ °C	[59, 62, 63]
Crystal form	Trigonal	[61, 64]
Melting enthalpy (ΔH_m)	142–155 J/g	[62, 65]
Density	1.21–1.34 g/cm ³	[66]
Tensile strength	80 Mpa	[59, 62, 65]
Young's modulus	8.6 GPa	[67]
Elongation	30%	[59, 67]
Hydrolytic degradation	PLA fibers 40% ~ 25 days	[63]
Viscosity	At room 1.258 g/cm ³ temperature	[66]
Permeability	Water 2954 ± 120 CO ₂ 32 ± 7 O ₂ 6 ± 0	[68]
Rockwell Hardness	~ 70–90	[67]

Insight into PLA degradation mechanism

PLA can be subjected to enzymatic, hydrolytic, microbial, and ultra-violet degradation (photodegradation). These processes are essential to determine the long-term impact of PLA-based materials. From a biomedical point of view, PLA degradation produces a decrease in

crystallinity percentage and molecular weight and generates smaller devices. The PLA's low degradation rate is not conducive to biomedical applications, especially bone tissue engineering. Practical strategies to address this drawback are grafting, copolymerization, compounding, and blending with other substances. It is very challenging to find a balance between degradation rates, mechanical strength, porosity, conformation, degree of crystallinity, shape, chemical cues, biodegradability, and biological activity to develop devices that ensure tissue's safe growth and sustain the biomaterial stability to perform the function for which it was implanted [69–71].

Modifications of PLA as a tissue engineering material

Decades ago, we could barely imagine the possibility of developing biomaterials using nanotechnology to improve humanity's quality of life. Today, biomaterials are widely used in endless applications, ranging from the food industry, textiles, and the medical and pharmacological industries, among others. A lot has been said regarding the applications of biomaterials in various areas of study or innovation. However, the field of biomaterials still has much to be explored. One of the multiple applications of biomaterials is the use of biopolymers as innovation materials in tissue engineering, for which the development of biomaterials focused on the treatment of

diseases has allowed the enhancement or improvement of cellular activity, serving as scaffolds for cell support or transport of pharmacological molecules that improve cell activity or viability allowing new treatments.

Additionally, an essential aspect of the development of biomaterials is the use of characteristics and functionalities selected from biological systems that provide us with essential information that can be utilized to develop new biomaterials that imitate the mentioned biological systems. For example, modified biopolymers mimic various aspects of the human body; according to Green *et al.* [72], hydrogels allow the creation of structures similar to the human body as scaffolds for *in vivo* tissue repair and for the cultivation of Stem cells. In addition to the above, the use of biopolymers in tissue engineering has characteristics of great interest, among which we can count on biocompatibility, bioactivity, non-toxicity, biodegradation, adaptable mechanical properties, and a biopolymer synthesis process of great convenience; all of these focused on the improvement of biological structures that mimic the body despite the use of synthetic polymers that although this might seem to generate incompatibility problems. The use of these natural or synthetic biopolymers allows adaptability to generate ideal materials that allow the functionality of the tissue, in addition to the fact that the area of tissue engineering and regenerative medicine emphasizes the use of these novel techniques for the essential goals of allowing the repair of damaged tissues or organs and that they continue to have origin from biofunctionality [73–78].

Considering the broad characteristics of materials, one question remains: What is needed to create an ideal tissue-engineered material? The cells' interactions with the materials must be considered to answer this question and how this interaction works to develop a tissue. One of the most popular characteristics is that the material has to be biocompatible with the cells and with the human body, which is essential to avoid immunogenic responses like allergies or rejection of the material from the body like a foreign body response, also with the compatibility, the toxicity is another relevant characteristic to have an ideal material. The polymer has to be safe for the organism and prove that the materials are not carcinogenic or have a potential risk of causing an illness. Also, biodegradability can be important too. A material that allows the regeneration of the tissue while it is disappearing can bring support to the cells and helps to promote the proliferation, migration, and cell growth in the scaffold. Cell interaction with the material is also needed to understand how the type of material and porosity influence the viability of the cells, or if the cells tend to create aggregates, it benefits the tissue development and how the cell adhesion works with a predetermined type of material. The 3D arrangement of the scaffolds influences cell differentiation and

the phenotype of the cells. Another last but not least important characteristic is the versatility of the polymer because it is advantageous to have materials that can be transformed into fibers, vesicles, or hydrogels to match biomedical applications [79–81] [Fig. 6].

As an emerging science, tissue engineering and regenerative medicine focus on developing materials that replace, restore or improve tissues or organs and enhance the cellular capacity to proliferate, migrate and differentiate into different cell types and specific tissues, respectively [82–84]. Therefore, the importance of these areas of the study lies in current therapies for various diseases ranging from the regeneration of skin, blood vessels, cartilage, heart tissue, bone, brain disorders, and even further in the urinary tract or gastrointestinal tract [85–91].

PLA as a tissue engineering material

Poly (lactic acid) has proven to be a biopolymer with great functionality in the biomedical area; considering the characteristics and properties of PLA that offer a large set of benefits and that combined can generate medical devices, we can highlight that within the properties of the biopolymer, the most important are biocompatibility, non-toxicity, and biodegradability [92].

Now, the central question is why the PLA needs to be modified. The copolymerization of PLA or the mixture of PLA with other polymers allows PLA to improve its properties and biological functions. According to Cheng *et al.* [93], the factors that influence the properties of polymers are the chemical components, the composition, and the morphological structure, among others. The authors also mention that they can improve the properties of both polymers. For example, the union between PLA and poly (glycolic acid) (PGA) results in a polymer with better properties such as low crystallinity and melting temperature (T_m), an example of an improved property is the copolymer poly (lactic glycolic acid) (PLGA), the concentration of the monomers can adjust the degradation of this biopolymer. Therefore, the modification of PLA allows that when grafting or making a mixture of other polymers with PLA, aspects of interest are modified according to the types of different polymers grafted to the base chain of PLA. In addition, a highly relevant benefit of PLA blends with other polymers is that when grafting or blending a new copolymer, it can be focused on a specific application that generates new research areas. Another benefit of PLA copolymerization is that the properties and characteristics, such as hydrophobicity of PLA, can be masked by other polymers and that mixture improves the capacity of PLA for more excellent compatibility [94]. However, it must be considered that some biopolymers are incompatible with PLA due to their hydrophobicity since, being an aliphatic compound, it tends to differ from hydrophilic compounds, leading to

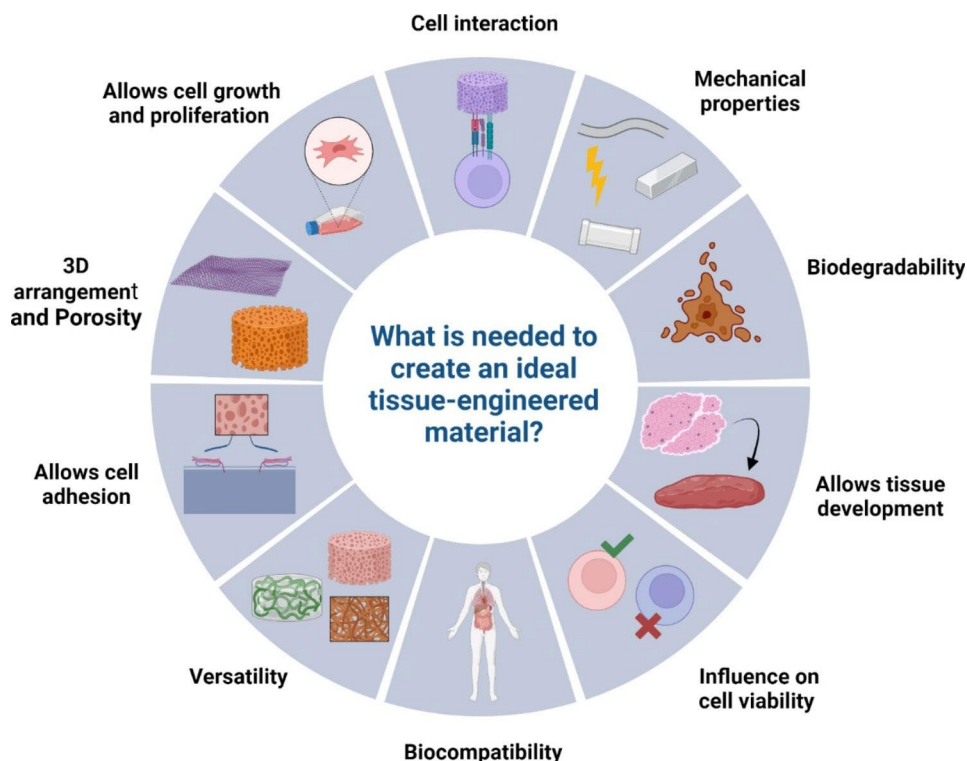


Fig. 6 Ideal tissue engineering material. Diagram of some relevant characteristics of an ideal tissue-engineered material for biomedical applications

low performance and inadequate performance response of the copolymer properties [95]. Likewise, chemical modifications are made to improve the crosslinking of the polymer, and this characteristic is relevant for synthesizing hydrogels since these modifications allow the crosslinking between polymers to be obtained with greater efficiency and ease [96].

Overview of some advances in PLA modifications: an insight into tissue engineering

According to the benefits of PLA copolymerization and its various properties, new biomaterials based on biopolymers have emerged due to the focus on improving treatments for various diseases.

PLA/Hydroxyapatite (HA)

One of the rising biomaterials developed is PLA modified with HA. HA has turned out to be a ceramic material with enormous potential for tissue engineering. It focuses on bone since the bone tissue in the crystalline phase is made up of HA in its natural state. In addition, ceramic materials such as HA have a significant porosity that allows this material to blend with bone tissue and bring oxygen. The biological importance of using HA in scaffolds is the natural presence in the bones. Therefore, HA increases the Ca^{2+} of the cell, thus allowing the proliferation of osteoblasts and promoting cell growth [97, 98].

On the other hand, PLA can be 3D printed for the design and synthesis of scaffolds that allow application in bone tissue engineering. Despite its outstanding biocompatibility, PLA has the disadvantage of not having suitable cell adhesion, which sometimes leads to the material being discarded for cell growth. However, this disadvantage can be overcome because when mixing HA, which is known to allow excellent cell adhesion and, therefore, cell proliferation, it allows cell adhesion by exerting electrostatic interactions with cell protein receptors; in this way, the capacities of the material would be balanced to be a candidate for the treatment of fractures or injuries where the bone can no longer regenerate itself [99, 100]. An exciting aspect worth mentioning is that for bone tissue engineering purposes, modifications to the surface of the scaffold can be made by adding molecules that improve cell adhesion.

One of the most versatile techniques that allow obtaining nanofibers with great ease and performance is electrospinning, which allows fibers to be created from a mixture of biopolymers in the order of nanometers to micrometers; this is attractive for the design of scaffolds. Another technique is air spray spinning, which covers areas with the compound of interest. This technique can help design and synthesize mats used as a scaffold for cell migration [101–104].

Currently, PLA polymeric fibers have been developed using the rotary jet spinning (RJS) method, which allows

generating effective wound dressing for skin tissue engineering. RJS provides several advantages, such as wound healing acceleration and improved cell proliferation, migration, and adhesion. Also, the resulting fibers offer adequate porosity and surface area while mimicking or imitating the extracellular matrix's structure well [105]. Recently, the RJS method permitted the production of antibacterial poly(ϵ -caprolactone)/Poly (lactic acid) fibers loaded with Vancomycin, which showed potential for use as a dressing intended for wound repair [106]. Furthermore, other research on newly developed PLA fibers combined the polymer with curcumin to produce a membrane by RJS for wound healing applications. The authors demonstrated that PLA / curcumin membranes are cyto-compatible with mouse embryonic fibroblasts [107]. It is worth mentioning the work highlighting the nature, wettability, morphology, and thermal properties of non-woven PLA produced from the RJS process. The quality and hydrophobic nature of the fibers were exhaustively assessed [108]. Besides, an exciting work reported the production via RJS of highly aligned and controlled nanofibers. The study described the dimensionless parameters used to prepare and scale up reliably nanofibers of several polymers, including PLA [109]. Lastly, it is of note a work employing centrifugal spinning for producing PLA / gelatin ultrafine fibers that were suitable for skin tissue engineering when loaded with ciprofloxacin [110]. Conclusively, the RJS method offers unique advantages and versatility to produce diverse types of fibers and scaffolds with controlled properties beneficial for tissue engineering.

On the other hand, carrier scaffolds of PLA combined with HA and poly (ethylene glycol) (PEG) have also been used for the transport of recombinant proteins that induce bone tissue formation, which allows an improvement in cell proliferation to repair the damaged tissue. Also, the use of erythropoietin as a precursor for proliferation and pleiotropic effects related to bone tissue [101, 111]. Even the use of PLA/HA coated with polypyrrole (PPy) has been reported, favoring cell proliferation due to its conductive properties [112]. Likewise, the PLA/HA copolymer has been used as a scaffold for regenerating dental pulp tissue that can help with treatments ranging from dental trauma to congenital disorders [113].

Another approach of the PLA/HA copolymer has been found for cartilage repair, where the biomaterial is used as a scaffold to carry recombinant proteins that will be precursors of cell proliferation of stem cells to differentiate into cartilage [114].

PLA/Poly (glycolic acid) (PGA)

The PLA/PGA copolymer has turned out to be a versatile biomaterial that has focused on the engineering of musculoskeletal tissues such as bone, menisci, or

cartilage because when trauma occurs, disease or congenital abnormalities exist. It is difficult to recover the tissue when it is damaged or lost; thus, using this material as a scaffold allows the replacement or regeneration of chondrocyte cells or osteoblasts. Likewise, it has been reported that PGA is an excellent biomaterial for cell growth, in addition to the fact that the mixture with PLA gives it greater effectiveness concerning cell growth and adhesion [115–119]. One of the characteristics that stand out and is convenient for the development of scaffolds for various tissues is the appropriate biocompatibility that this copolymer has and in addition to the fact that it can be designed in different structures such as scaffolds or nanocomposites for various biomedical applications such as, in the regeneration of trachea, spinal cord, and the brain. The latter case is of great relevance in the current biomedical area since being able to develop biomaterials that allow the regeneration of damaged brain tissues is a premise for neurodegenerative diseases using scaffolds with Schwann cells and neural Stem cells. Unlike other applications for this copolymer, the mechanical strength is not essential, but the biodegradation rate is determined by the polymer's crystallinity and molecular weight. According to Agrawal *et al.* [120], the PLA/PGA copolymer degradation rate depends on the exact rate of PLA and PGA monomer contained in the polymer, and this biodegradation is conducted by breaking ester bonds hydrolytically. Likewise, the modifications of the topology of the copolymer open an opportunity to obtain more excellent cell adhesion and, thus, better cell proliferation and differentiation [121].

On the other hand, another relevant application is the regeneration of bone tissue for facial reconstruction, a common injury caused by trauma [122]. Another function of the scaffolds of this copolymer is focused on the regeneration of cardiac tissue, which is a well-known heart disease and is a latent risk today. Therefore, the focus on cardiac tissue regeneration is vital for current research. Tissue engineering allows the development of biografts with skeletal myoblast and endothelial cells, among others, to produce a scaffold that carries the cells allowing correct oxygenation, adhesion, cell proliferation, and angiogenesis, as in the scaffolds for neuronal tissue. The rate of degradation in cardiac tissues is essential and has adequate *in vivo* prevalence to migrate the cells, and the proliferation process occurs adequately [123, 124]. A last point of interest is the use of PLA/PGA nanofiber scaffolds with drugs for their administration, and an example is the use of PLA/PGA/Ibuprofen nanofibers for administration in chronic wounds and to help relieve pain caused by these diseases. These scaffolds adhere to cells and release drug-producing benefits such as pain reduction and relief of wound inflammation [125].

PLA/Poly(butylene-adipate-Co-terephthalate) (PBAT)

There is a wide range of applications for the mixture of PLA and PBAT polymers, mainly using electrospinning techniques for the generation of nanofibers of these compounds, in which different modifications can be made to parameters such as the types of solvents, mixing ratio of binary solvents, polymer blend concentration, polymer blend ratio, among others [126, 127]. This combination of compounds has also been used in other areas of study, for example, for the production of films, resulting in better tensile strength and significantly improving the joint use of both compounds than in isolation [128]. On the other hand, these polymers have been used to manufacture biomembranes and as materials in food packaging, resulting in a high degree of versatility and a wide range of applications [129].

Other investigations have shown that both compounds have excellent biocompatibility, resulting in promising materials used in bone tissue engineering [130]. This mixture has also favored the proliferation of fibroblasts since it has accessible mechanical properties, high porosity, well-connected microporous structures, excellent water permeability, and good biocompatibility to support the formation of new tissues, allowing these cells to maintain their phenotypic shape. As the PBAT content increased, the mean diameter of the PLA/PBAT scaffolds decreased while the mechanical properties improved [131, 132].

PLA/PEG

Even though there are many ways of designing biomaterials, hydrogels allow a favorable environment to be cell-carrying scaffolds. PEG has the characteristic of improving the biocompatibility, hydrophilicity, ductility, and flexibility of the copolymers. However, PLA is a polymer with low hydrophilicity; in addition to being molded in different structures, they have the characteristic of having significant porosity, which is of great help for cell proliferation and oxygenation, and this, in turn, is of great interest for bone tissue engineering because it forms a versatile matrix to serve as a carrier scaffold for cells or recombinant proteins [133–135]. Taking advantage of PEG's good miscibility with organic solvents and its biodegradation by hydrolysis, and that PEG surfaces can be modified, they can be used to develop micelles that can be drug vehicles or function as materials with antibacterial activity. According to Tessmar et al. [136], an attractive property of PEG is that because it is an uncharged molecule, it tends to form highly hydrated polymer coils on the copolymer surfaces, and most importantly, it can repel proteins. This property is used because, with modifications, it is possible to obtain micelles or other biomaterials with specific interactions resulting from peptide sequences grafted to the copolymer.

In addition, when PLA/PEG are copolymerized, they have the characteristic of improving PLA degradation properties, biocompatibility, non-toxicity, and good solubility, which allows scaffolds to be designed with good porosity, resistance, and degradation. Applying the copolymer to wound healing is the implementation of platelet growth factors to induce cell proliferation and thus have a better regenerative process [137–139].

Regarding the application of bone tissue engineering, the porosity property of PLA/PEG material has allowed the development of three-dimensionally printed scaffolds. Also, the use of techniques such as electrospinning that allows the creation of nanofibers for the design of mats that are of great support for the synthesis of scaffolds where cells can have excellent adhesion, proliferation, migration, and nutrition of Stem cells since it has been reported that Stem cells have had a more significant differentiation in their potential because they favor the expression of osteogenic cell markers [140–142].

PLA/Lignin

Cellulose is not only one of the most critical polysaccharides, but also lignin is one of the most abundant polysaccharides all over the world. Because of the above, lignin has excellent potential to be used as a polymer in multiple biomedical applications since this polysaccharide has properties of great interest, such as antimicrobial, antioxidant, anti-ultra-violet (UV), biocompatibility, and non-toxicity properties [143].

Within the various applications for the PLA/Lignin copolymer, the use of scaffolds made from nanofibers by electrospinning focused on cartilage and bone tissue engineering is one of the leading applications today [144–146]. In addition to the fact that the PLA/Lignin copolymer has the characteristic of enhancing the properties of mechanical, thermal, and UV resistance and provides it with the essential characteristic of improving resistance to oxidative stress. The contributions of lignin enhance the properties and add new ones with which PLA alone does not count. This fact makes the copolymer attractive for the development of biomaterials, such as the development of PLA/Lignin films through physical methods like the mixture of these two polymers for various applications in the biomedical area with the use of Stem cells [147].

A novel application of the PLA/Lignin copolymer is the use of PGA to make a nanoparticle that has drug delivery functionality with a specialized focus on those therapies that are difficult to administer or have a beneficial result for patients; an example is the use of these nanoparticles to improve drug delivery efficiency for patients with triple-negative breast cancer. In this example, lignin improves the drug delivery system, and the copolymer improves cell non-toxicity and the biocompatibility of

the nanoparticle, in addition to the fact that thanks to its compact size, it makes it a useful resource for drug delivery by nanoparticles [148]. In addition to the fact that a property of lignin that is of foremost importance is that it can form porous materials, and this is of great interest due to its multiple applications in tissue engineering as a common element for the creation of scaffolds for the regeneration of damaged tissue [149].

PLA/Poly(pyrrole) (PPy)

One of the most promising materials due to its high conductive properties is PPy, an inorganic polymer. Polypyrrole has been considered one of the most useful in studying neuronal regeneration. Due to this, tissue engineering focused on neuronal tissues has used PPy as a polymer in combination with various polymers to develop scaffolds. One of these exciting materials is PLA. Together with PPy, they make up one of the most valuable materials because PLA is a highly biocompatible polymer with the organism and has high degradation capacities in conjunction with PP and PLA, which gives it these characteristics, enhancing biocompatibility. Something of considerable interest for neural tissue engineering is the ability of the material to electrically stimulate the proliferation, adhesion, and cell growth of damaged neuronal tissue. However, it is also of great interest for other tissues with electroactive potentials, such as the heart [150].

Among the applications of the copolymer is the creation of nanofibers by electrospinning of PLA/PPy, according to Tian *et al.* [151] report that the copolymer nanofibers were found to have better adhesion, viability, and cell proliferation. Then, the development of PLA/PPy nanofiber scaffolds to support bone marrow stem cells with the premise that cell regeneration is induced in spinal cord injuries. It is worth mentioning that these studies have been conducted *in vivo* in Wistar rats [152]. Another application has been made with the development of conductive fibers with conductive centers of PLA/PPy. Surface modifications were made to these fibers by adding proteins to make a bioactive scaffold that allows better adhesion and biocompatibility while preserving electroactive properties. These properties are of great interest for repairing damaged tissues, in addition to the fact that these scaffolds can be electrically stimulated, which allows the support of cell adhesion [153, 154].

PLA/Chitosan (CHI)

Chitosan is a natural polymer made from renewable resources obtained from waste from the fishing industry and the shell of mollusks. It is a biocompatible, biodegradable material with antibacterial activity and allows wound healing [155, 156]. Additionally, chitosan and its

derivatives are promising candidates to serve as support material in tissue engineering applications because of their characteristics of porous structure, gel-forming properties, ease of chemical modification, and high affinity with macromolecules *in vivo*, among others [157].

In tissue engineering, there has been a more remarkable boom in the study of the regeneration of damaged tissues as an alternative to autografts, where the mixture of PLA and chitosan stands out [158]. Nowadays, a great variety of natural and biocompatible compounds have been used for the elaboration of scaffolds with applications in regenerative medicine, chitosan being the material of choice since scaffolds present antibacterial and proangiogenic activity and mimic the extracellular matrix [156, 159]. In addition, one of the main limitations of PLA is its low cell affinity because it has poor cell recognition sites and low hydrophilicity; for which chitosan has allowed us to overcome these limitations of the scaffolds of PLA since it has minimal reaction to foreign bodies and good hydrophilicity, has been used as a surface modification material to improve cell attachment and proliferation in scaffolds [160]. Another advantage of the mixture of PLA and chitosan polymers is that it reduces acid by-products which can cause inflammatory reactions in the tissues and generate clinical failures; the above were present in the scaffolds that were only composed of PLA [161]. One of the most used techniques for producing PLA and chitosan nanofibers is electrospinning, and it is intended that these spun nanofibers be used in the native extracellular matrix for tissue engineering [162].

An example of the use of these scaffolds made by electrospun emulsion is in the regeneration of bone tissue, which turned out to be compatible with cells and biodegradable for periodontal bone regeneration by regulating their mechanical and biological properties; chitosan also promoted cell adhesion and osteogenic differentiation of bone marrow stem cells (BMSCs) [163]. Another example of its use in the regeneration of bone tissue is in preparing a tunable biomimetic matrix composed of chitosan, which promotes osteoconduction, and positively affects the behavior of osteoblasts [164]. In the same way, multiple applications of scaffolds composed of PLA and chitosan fibers have been found for cardiac tissue engineering and to accelerate myocardial regeneration since, in a particular proportion, they support the viability of cardiomyocytes, cause cell elongation and enhance the production of sarcomeric α -actinin and troponin I. On the other hand, PLA and chitosan nanofibers have also been used for the treatment of cuteness injuries caused by burns. These three-dimensional scaffolds were made from electrospinning techniques and are not toxic to skin cells, and can mimic the extracellular matrix, mainly composed of nanofibrous proteins [165].

PLA / Poly (Caprolactone) (PCL)

One of the mixtures with the most significant potential in tissue engineering is the one conducted with PLA and PCL. These polymeric compounds have allowed the realization of scaffolds with multiple applications in regenerative medicine; their main advantages are their high purity, adequate processing, and excellent mechanical properties. Likewise, it should be noted that they are biodegradable materials whose degradation products can be reabsorbed.

Lactic acid presents various physiological and metabolic pathways from being a small organic molecule. The addition of these two compounds enhances the biomechanical performance of the constructions, and some studies have shown improvements in their mechanical and biological properties [166, 167].

Among the primary uses of these materials is the construction of bone scaffolds using an indirect 3D printing approach, in which the cells of interest became viable and proliferated, as well as increased biocompatibility and osteoinduction properties [168]. Another application of this mixture is in blood vessel tissue engineering, where scaffolds have been developed that mimic their architecture through sequential electrospinning technologies, resulting in excellent candidate scaffolds for this area. In the same way, its enormous potential for application in the field of vascular patches has been demonstrated [169, 170]. Some research indicates that the expression levels of elastin, angiopoietin, laminin-4 α and -5 α increased in PCL and PLA nanofibers without any exogenous factor, in addition to the fact that they are significantly less hydrophobic and have less resistance to traction [171]. Given the above, it should be noted that the high biocompatibility between both compounds and their physical properties make them a suitable material for the replacement of blood vessels, which in the future would allow the possibility of functionalizing that material with a variety of molecules and modulate inflammatory and coagulative responses. Then, suitable devices would be obtained to replace native vessels [172, 173]. Other examples of composite grafts exhibited significant improvements in mechanical characteristics compared to single-material devices, particularly in compression and torsional strength, which are common problems with single-polymer vascular grafts compared to composite vascular grafts [174].

In addition to its use in the regeneration of vascular tissue, the mixture of PLA and PCL polymers has made it possible to develop viable nerve tissue substitutes by combining scaffolds with transplanted cells and growth factors. Inkjet technology is attractive for manufacturing these scaffolds due to the incorporation of non-contact approaches that allow precise volumes of material to be deposited with high speed and precision at destination

sites [175]. Figure 7 summarizes the previous section in a graph showing the most common PLA blends and their applications. Also, Table 3 summarizes the PLA combinations mentioned above, highlighting advantages and some relevant properties.

The emphasis on recent advances in PLA derivatives

Recently, several works have highlighted the most innovative developments for PLA derivatives. Ren et al. [205] reported a bimodal cell structure PLA/ cellulose nanocomposite synthesized by depressurization foaming as a possible thermal insulation prospect material. Another significant innovation is the production of chitosan/collagen hydrogel scaffolds from 3D printed PLA strut and cellulose nano-fibers intended for use in cartilage tissue engineering. The composite showed no cytotoxic effect on mesenchymal stem cells and enabled cell growth, attachment, proliferation, and migration through the scaffolds [206]. Also noted is the use of a freeze-drying technique to prepare a PCL/PLA scaffold containing zirconium (n-ZrO₂) nanoparticles. The scaffolds were subsequently coated with polypyrrole and then enhanced their hydrophilicity and supported in vitro human corneal epithelial cell viability, attachment, and proliferation, suggesting a possible use in regenerative medicine [207]. Lastly, Ye et al. called attention to the PCL/ PLA/ microcrystalline cellulose composites fabricated via extrusion technology. The innovative development exhibited high biocompatibility and adhesion of human breast cancer cells, indicating a bright future in bioengineering research [208].

On the other hand, some current research themes regarding PLA-based materials are the shape memory effect, the piezoelectric properties, and the injectability. The shape memory effect is the polymer's ability to change from the initial shape to a stress-free form. The latter state maintains the shape recovery until it is triggered externally with a stimulus. 4D printing implies changing the functionality or structural property of PLA-based biomaterial tridimensionally printed [209]. Polybutylene succinate/PLA composite filament was prepared by 4D printing, and the scaffold showed potential for use in tissue engineering [210]. Maleic anhydride grafted onto PLA was used as a compatibilizer (2 wt%) on the shape memory abilities of poly(ethylene glycol)/ PLA blends, allowing for the biomaterials to be optimized for usability as scaffolds with improved chain entanglement and interfacial adhesion [211].

In recent years, researchers have paid much attention to the electroconductive and piezoelectric properties and behavior of PLA-based materials for their potential to mimic bone tissues. Ferroelectrets films created from PLA can be used to prepare biosensors involved in the development and growth of cells [212]. A biphasic

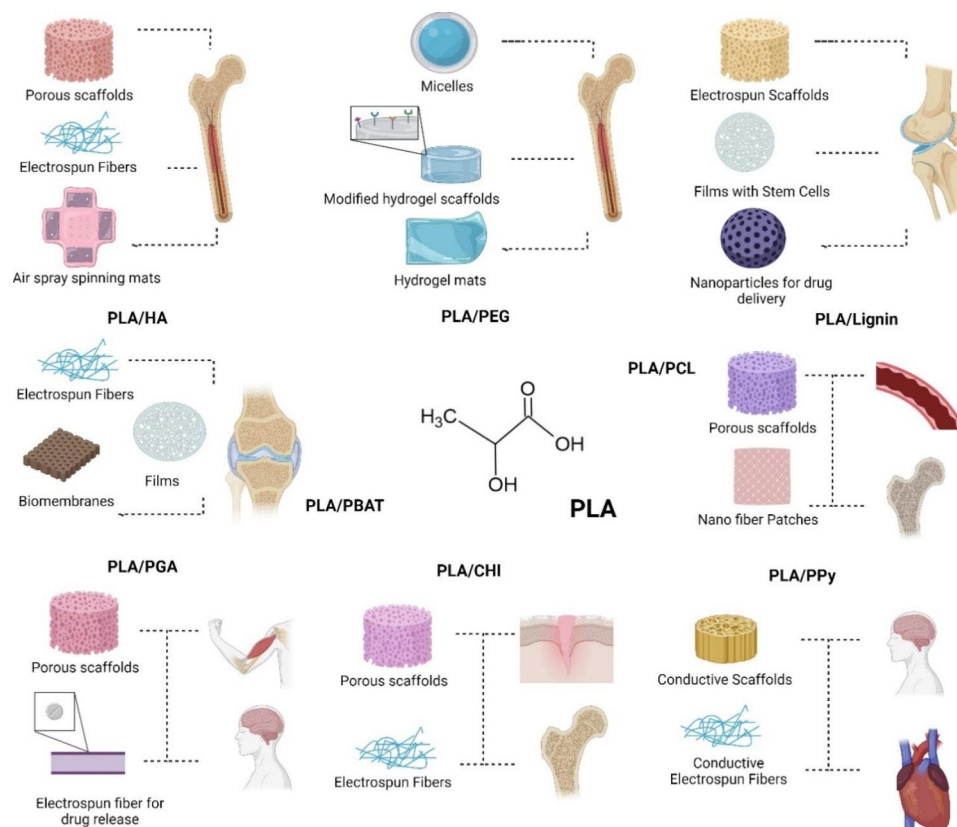


Fig. 7 Poly (lactic acid) modifications and applications

layered structure was synthesized from electrospun mats of piezoelectric polyvinylidene fluoride/PLA embedded in a terpolymer hydrogel of alginate/gelatin/polypyrrole-grafted-gelatin to form osteon-mimetic samples [213]. Biopiezoelectric materials, such as barium titanate/PLA composites, mimicked the microenvironment of bone tissue successfully and guided it through regeneration [214]. Concerning the injectability of PLA-based polymers, the scientific community has found that these biomaterials can act as an aqueous reservoir to treat defect shapes in bone tissue engineering because they can be molded easily and exhibits good biodegradability, rheology, selectivity, and targeting capability [212, 215–217].

Figure 8 represents the performance of PLA-based materials added with antimicrobials for *in vitro* and *in vivo* research. A skin biopsy provides autologous dermal fibroblast that can be combined with PLA copolymers after cell culture and expansion to produce wound dressings. The design of new types of PLA derivatives architecture supplies the necessary micro-environment for wound healing [218, 219].

Some of the most recent and exciting works found in the literature are the *in vitro* increase of the bioactivity and interface strength of PLA hybrid coatings which was reinforced with hydroxyapatite – Al_2O_3 [220]; the inhibitory capacity against *S. aureus*. and *E. coli*, the

antioxidant capacity, and the interphase compatibility were improved *in vitro* for novel poly(butylene succinate)/PLA blend combined with glycyrrhetic and rosmarinic acids as compared to neat PLA [221]; and the *in vitro* and *in vivo* study of the anti-oxidative and anti-fungal efficacy of PLA fibers containing *Ocimum gratissimum* L. and *Ocimum basilicum* L. against *Aspergillus niger* and *Aspergillus carbonarius* [222].

Limitations of the PLA as a biomaterial

PLA is a versatile polymer with many applications in various areas. Despite this, working with PLA can have disadvantages that must be considered since being a semi-synthetic polymer; it can have problems such as the lack of recognition of cell signaling, lack of adhesion, or even hydrophobicity that affects tissue development *in vivo* depending on the applications in which it is involved. Not to mention that the same advantage provided by the degradability of PLA can be a disadvantage since, being volatile and prone to hydrolysis, it may not be viable for specific biomedical applications [223, 224].

Moreover, it is expensive compared to petrochemicals, with a more intense manufacturing method and a lower yield than conventional polymers. The PLA working temperature is pretty low, and the co-blending is challenging to implement.

Table 3 Summary of the combinations of PLA

	Modes of fabrication	Relevant properties	Advantages	Disadvantages	Refs.
PLA/HA	3D printing Electrospinning Air spray/jet spinning	Biocompatibility Porosity Versatility	Bone in crystalline phase is made by HA Allow blending with the bone Increases the Ca ²⁺ in the cell Allow the proliferation of osteoblasts and promote cell growth Can be used as carrier scaffolds to transport proteins	Poor cell adhesion of PLA at its own Discarded for cell growth	[176–181]
PLA/PGA	Biografts Electrospinning	Biocompatibility Versatility Biodegradation rate depends on molecular weight	PLA/PGA is an excellent material for cell growth Allows osteoblast regeneration PLA/PGA mixture is convenient for cell adhesion Topology modifications enhance cell proliferation, adhesion, and differentiation	Fast degradation rate Risk of inflammation The process of cross-linking in hydrogels sometimes is not effective	[118, 182–185]
PLA/PBAT	Electrospinning Biomembranes	Tensile strength Versatility Biocompatibility Accessible mechanical properties Porosity Water permeability Interconnected microporous	Wide range of applications Favor the proliferation rate	The mean diameter of the PLA/PBAT scaffolds decreased while the mechanical properties improved	[129, 186, 187]
PLA/PEG	Micelles 3D printing Electrospinning	Hydrophilicity Ductility Flexibility Porosity Versatility Biodegradation by hydrolysis Can repel protein	PEG improves the biocompatibility of the copolymers with which it is mixed Help for cell proliferation and oxygenation PEG has good miscibility with organic solvents	Poor cell adhesion	[185, 188–192]
PLA/Lignin	Electrospinning Nanoparticles	Antimicrobial Antioxidant Anti-ultra-violet (UV) Biocompatibility Non-toxicity Porosity	Enhance the mechanical properties of the copolymers	The use of high concentrations of sodium chloride used as a solvent cause phase separation	[143, 190, 193–196]
PLA/PPy	Electrospinning Hydrogels	Conductivity Biocompatibility Biodegradation	Electrically stimulate the proliferation, adhesion, and cell growth in potential electroactive tissues	Low solubility The PPy tends to be fragile	[192, 197, 198]
PLA/Chi	Electrospinning Biomembranes Micelles Hydrogels Nanoparticles	Biocompatible Biodegradable Antibacterial activity Porosity Gel-forming properties High affinity with macromolecules	It is a natural polymer made from renewable sources Allows wound healing	It has poor cell recognition sites and low hydrophilicity Poor mechanical properties	[162, 196, 199–201]
PLA/PCL	Electrospinning Biomembranes Inkjet technology 3D printing	High purity Adequate processing Excellent mechanical properties Biocompatibility Biodegradation	The expression levels of elastin, angiotensin, laminin-4a and –5a increased in PCL and PLA nanofibers without any exogenous factor	Degradation products can be reabsorbed PLA/PCL are less hydrophobic and have less resistance to traction	[168, 199, 202–204]

PLA is known for its controllable degradation rate and non-toxic components of degradative products. However, PLA-based degradation behavior depends on the molecular weight and glass transition temperature. Therefore, a feasible procedure must be attained to avoid undesirable degradation products towards *in vitro* and

in vivo performance. Acid degradation by-products may produce inflammatory reactions and low cell affinity. It is thus crucial to prepare new added-value PLA biomaterials considering these issues.

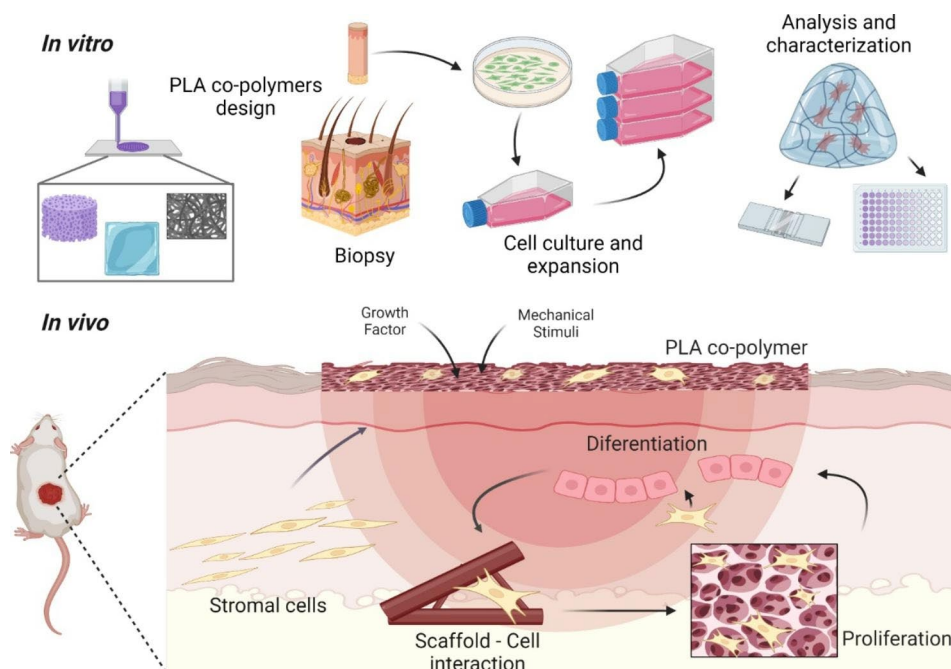


Fig. 8 PLA and PLA-based materials designed for in vitro and in vivo performance

Conclusion

The capability of the human being to regenerate damaged tissues by itself is limited. Today diseases are fought and studied differently. Therefore, it becomes more complex to determine an effective treatment that is effective, efficient, and highly accurate. In addition, treatments must be friendly to the environment and low cost. Thus, PLA is an attractive biopolymer due to its multiple properties (biocompatibility, biodegradation, mechanical strength, non-toxicity) that benefit when developing new treatments within tissue engineering. It should be noted that these properties can be enhanced according to the modifications that can be made to PLA when mixed with other polymers, which opens up a broad spectrum of possibilities for the potential treatment of diseases.

Conclusively, the current trends in PLA-based research that are influencing the future of tissue engineering applications are the evolution of conventional electrospinning to needleless electrospinning, such as ultrasound-enhanced electrospinning, edge electrospinning (jet spinning), and near-field electrospinning to produce high-quality nanofibers scale-up; the scope widening by the synthesis of new tailored devices in the form of nano- and micro- capsules, particles, and hydrogels; the preparation of cutting-edge membranes for skin tissue engineering; the advances in 3D and 4D printing technologies; and the versatility in forming new types of complex composites and scaffoldings. This review is a good roadmap for implementing new approaches to strengthen the biomedical fields through the invention of feasible medical surgeries, derma, membranes covering, cosmetics,

tissue engineering, and scaffolding. It is also worth mentioning that PLA represents a carbon emissions reduction, another point in its favor, while multiple research opportunities are waiting to be discovered.

Acknowledgements

Samanta Castañeda Rodríguez is currently an MSc student in the "Maestría en biomedicina y biotecnología molecular" Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, and received a fellowship from CONACYT, Mexico. Maykel González-Torres acknowledges the financial support by INVESTIGADORES POR MEXICO DE CONACYT. The authors thank www.Biorender.com for the figures created.

Authors' contributions

SCR, MGT, RMRA, MLDPA, GLG, E.S.G., and JS-R, made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis, and interpretation, or in all these areas—that is, revising or critically reviewing the article; giving final approval of the version to be published; agreeing on the journal to which the article has been submitted; and confirming to be accountable for all aspects of the work. All authors have read and approved the final manuscript.

Funding

No funding was received.

Availability of data and material

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors wish to confirm that there are no known conflicts of interest associated with this publication, and there has been no significant financial support for this work that could have influenced its outcome.

Received: 21 November 2022 / Accepted: 7 March 2023

Published online: 20 March 2023

References

- Ulery BD, Nair LS, Laurencin CT. Biomedical applications of biodegradable polymers. *J Polym Sci Part B: Polym Phys*. 2011;49(12):832–64.
- Kumar SSD, Abrahamse H. Advancement of nanobiomaterials to deliver natural compounds for tissue engineering applications. *Int J Mol Sci*. 2020;21(18):6752.
- Sin L, Rahmat A, Rahman W. Degradation and stability of poly (lactic acid). *Polylactic Acid*. 2013:247 – 99.
- Carvalho TSS, Ribeiro N, Torres PMC, Almeida JC, Belo H, Araújo J. Magnetic poly(lactic acid)-calcium phosphate-based biocomposite as a potential biomaterial for tissue engineering applications. *Mater Chem Phys*. 2023;296:127175. <https://doi.org/10.1016/j.matchemphys.2022.127175>.
- Maleki H, Azimi B, Ismaeilmoghadam S, Danti S. Poly(lactic acid)-Based Electrospun Fibrous Structures for Biomedical Applications. *Appl Sci*. 2022;12(6):3192.
- Miranda CC, Gomes MR, Moço M, Cabral JMS, Ferreira FC, Sanjuan-Alberte P. A concise review on Electrospun scaffolds for kidney tissue Engineering. *Bioeng (Basel)*. 2022;9(10). <https://doi.org/10.3390/bioengineering9100554>.
- Milovanovic S, Pajnik J, Lujic I. Tailoring of advanced poly(lactic acid)-based materials: a review. *J Appl Polym Sci*. 2022;139(12):51839. <https://doi.org/10.1002/app.51839>.
- Vlachopoulos A, Karlioti G, Balla E, Daniilidis V, Kalamas T, Stefanidou M, et al. Poly(lactic Acid)-Based microparticles for drug delivery applications: an overview of recent advances. *Pharmaceutics*. 2022;14(2). <https://doi.org/10.3390/pharmaceutics14020359>.
- Ilyas RA, Zuhri MYM, Aisyah HA, Asyraf MRM, Hassan SA, Zainudin ES, et al. Natural Fiber-Reinforced Poly(lactic Acid), Poly(lactic Acid) Blends and their composites for Advanced Applications. *Polym (Basel)*. 2022;14(1). <https://doi.org/10.3390/polym14010202>.
- Joseph TM, Kallingal A, Suresh AM, Mahapatra DK, Hasanin MS, Haponiuk J et al. 3D printing of poly(lactic acid): recent advances and opportunities. *Int J Adv Manuf Technol*. 2023:1–21. doi: <https://doi.org/10.1007/s00170-022-10795-y>.
- More N, Avhad M, Utekar S, More A. Poly(lactic acid) (PLA) membrane—significance, synthesis, and applications: a review. *Polym Bull*. 2023;80(2):1117–53. <https://doi.org/10.1007/s00289-022-04135-z>.
- Grigora M-E, Terzopoulou Z, Baciu D, Steriotes T, Charalambopoulou G, Gounari E, et al. 3D printed poly(lactic acid)-based nanocomposite scaffolds with bioactive coatings for tissue engineering applications. *J Mater Sci*. 2023;58(6):2740–63. <https://doi.org/10.1007/s10853-023-08149-4>.
- Martinez FAC, Balciunas EM, Salgado JM, González JMD, Converti A, de Souza Oliveira RP. Lactic acid properties, applications and production: a review. *Trends Food Sci Technol*. 2013;30(1):70–83.
- Benninga H. A history of lactic acid making: a chapter in the history of biotechnology. Springer Science & Business Media; 1990.
- Dorgan J, Braun B, Wegner J, Knauss D. Poly (lactic acids): a brief review. 2006.
- Carothers WH, Dorough G, Natta Fv. Studies of polymerization and ring formation. X. The reversible polymerization of six-membered cyclic esters. *J Am Chem Soc*. 1932;54(2):761–72.
- Kulkarni R, Pani K, Neuman C, Leonard F. Poly(lactic acid) for surgical implants. *Arch Surg*. 1966;93(5):839–43.
- Huang S, Xue Y, Yu B, Wang L, Zhou C, Ma Y. A review of the recent developments in the bioproduction of poly(lactic acid) and its precursors optically pure lactic acids. *Molecules*. 2021;26(21):6446.
- Södergård A, Stolt M. Properties of lactic acid based polymers and their correlation with composition. *Prog Polym Sci*. 2002;27(6):1123–63.
- Idler C, Venus J, Kamm B. Microorganisms for the production of lactic acid and organic lactates. *Microorganisms in biorefineries*. 2015:225 – 73.
- Yankov D. Fermentative Lactic Acid Production From Lignocellulosic Feedstocks: From Source to Purified Product. *Frontiers in Chemistry*. 2022;10.
- Wang Y, Wu J, Lv M, Shao Z, Hungwe M, Wang J et al. Metabolism characteristics of lactic acid bacteria and the expanding applications in food industry. *Frontiers in bioengineering and biotechnology*. 2021:378.
- Klotz S, Kaufmann N, Kuenz A, Prübe U. Biotechnological production of enantiomerically pure d-lactic acid. *Appl Microbiol Biotechnol*. 2016;100(22):9423–37.
- Huang LP, Jin B, Lant P, Zhou J. Biotechnological production of lactic acid integrated with potato wastewater treatment by *Rhizopus arrhizus*. *J Chem Technol Biotechnology: Int Res Process Environ Clean Technol*. 2003;78(8):899–906.
- Hagen R. PLA (Polylactic Acid). Reference Module in materials Science and Materials Engineering. Elsevier; 2016.
- Huang LP, Dong T, Chen JW, Li N. Biotechnological production of lactic acid integrated with fishmeal wastewater treatment by *Rhizopus oryzae*. *Bioprocess Biosyst Eng*. 2007;30(2):135–40.
- Zhang ZY, Jin B, Kelly JM. Production of lactic acid from renewable materials by *Rhizopus fungi*. *Biochem Eng J*. 2007;35(3):251–63.
- Petrova P, Petrov K. Lactic acid fermentation of cereals and pseudocereals: ancient nutritional biotechnologies with modern applications. *Nutrients*. 2020;12(4):1118.
- Pielech-Przybylska K, Balcerek M, Ciepeliowski G, Pacholczyk-Sienicka B, Albrecht Ł, Dziekońska-Kubczak U, et al. Effect of co-inoculation with *Saccharomyces cerevisiae* and lactic acid bacteria on the content of propan-2-ol, acetaldehyde and weak acids in fermented distillery mashes. *Int J Mol Sci*. 2019;20(7):1659.
- Peetermans A, Foulquié-Moreno MR, Thevelein JM. Mechanisms underlying lactic acid tolerance and its influence on lactic acid production in *Saccharomyces cerevisiae*. *Microb Cell*. 2021;8(6):111–30.
- Ilmén M, Koivuranta K, Ruohonen L, Rajgarhia V, Suominen P, Penttilä M. Production of L-lactic acid by the yeast *Candida sonorensis* expressing heterologous bacterial and fungal lactate dehydrogenases. *Microb Cell Fact*. 2013;12(1):1–15.
- Esquivel-Hernández DA, Pennacchio A, Torres-Acosta MA, Parra-Saldívar R, de Souza Vandenberghe LP, Faraco V. Multi-product biorefinery from *Arthrospira platensis* biomass as feedstock for bioethanol and lactic acid production. *Sci Rep*. 2021;11(1):1–15.
- Angermayr SA, van der Woude AD, Correddu D, Kern R, Hagemann M, Hellingwerf KJ. Chirality matters: synthesis and consumption of the d-enantiomer of lactic acid by *Synechocystis* sp. strain PCC6803. *Appl Environ Microbiol*. 2016;82(4):1295–304.
- Angermayr SA. Synthetic biology of cyanobacterial cell factories. *Universiteit van Amsterdam [Host]*; 2014.
- Wijffels RH, Kruse O, Hellingwerf KJ. Potential of industrial biotechnology with cyanobacteria and eukaryotic microalgae. *Curr Opin Biotechnol*. 2013;24(3):405–13.
- Liu L, Pohnert G, Wei D. Extracellular metabolites from industrial microalgae and their biotechnological potential. *Mar Drugs*. 2016;14(10):191.
- Carr FJ, Chill D, Maida N. The lactic acid bacteria: a literature survey. *Crit Rev Microbiol*. 2002;28(4):281–370.
- Nakata K, Miyazaki N, Yamaguchi H, Hirose M, Kashiwagi T, Kutumbarao NH, et al. High-resolution structure of phosphoketolase from *Bifidobacterium longum* determined by cryo-EM single-particle analysis. *J Struct Biol*. 2022;214(2):107842.
- Kandler O. Carbohydrate metabolism in lactic acid bacteria. *Antonie Van Leeuwenhoek*. 1983;49(3):209–24.
- Wood BJ, Holzappel W. The genera of lactic acid bacteria. Springer Science & Business Media; 1992.
- Vinderola G, Ouwehand A, Salminen S, von Wright A. Lactic acid bacteria: microbiological and functional aspects. *Crc Press*; 2019.
- Porto MCW, Kuniyoshi TM, Azevedo P, Vitolo M, Oliveira RS. *Pediococcus* spp.: an important genus of lactic acid bacteria and pediocin producers. *Biotechnol Adv*. 2017;35(3):361–74.
- Zuniga M, Pardo I, Ferrer S. An improved medium for distinguishing between homofermentative and heterofermentative lactic acid bacteria. *Int J Food Microbiol*. 1993;18(1):37–42.
- Garvie EI. Bacterial lactate dehydrogenases. *Microbiol Rev*. 1980;44(1):106–39.
- Von Wright A, Axelsson L. Lactic acid bacteria: an introduction. *Lactic acid bacteria*. CRC Press; 2019. pp. 1–16.
- Abedi E, Hashemi SMB. Lactic acid production—producing microorganisms and substrates sources-state of art. *Heliyon*. 2020;6(10):e04974.

47. Wang L, Cai Y, Zhu L, Guo H, Yu B. Major role of NAD-dependent lactate dehydrogenases in the production of L-lactic acid with high optical purity by the thermophile *Bacillus coagulans*. *Appl Environ Microbiol*. 2014;80(23):7134–41.
48. Lund J, Aas V, Tingstad RH, Van Hees A, Nikolić N. Utilization of lactic acid in human myotubes and interplay with glucose and fatty acid metabolism. *Sci Rep*. 2018;8(1):1–14.
49. Abdel-Rahman MA, Tashiro Y, Sonomoto K. Recent advances in lactic acid production by microbial fermentation processes. *Biotechnol Adv*. 2013;31(6):877–902.
50. Henard CA, Freed EF, Guarnieri MT. Phosphoketolase pathway engineering for carbon-efficient biocatalysis. *Curr Opin Biotechnol*. 2015;36:183–8.
51. Mokoena MP. Lactic acid bacteria and their bacteriocins: classification, biosynthesis and applications against uropathogens: a mini-review. *Molecules*. 2017;22(8):1255.
52. Wisselink H, Weusthuis R, Eggink G, Hugenholtz J, Grobber G. Mannitol production by lactic acid bacteria: a review. *Int Dairy J*. 2002;12(2–3):151–61.
53. Salvetti E, Fondi M, Fani R, Torriani S, Felis GE. Evolution of lactic acid bacteria in the order Lactobacillales as depicted by analysis of glycolysis and pentose phosphate pathways. *Syst Appl Microbiol*. 2013;36(5):291–305.
54. Berry M, Symposium. *Advances in Liver Pathology [Abridged] The Liver and Lactic Acidosis*. SAGE Publications; 1967.
55. Foucher CD, Tubben RE. Lactic acidosis. *StatPearls [Internet]*. StatPearls Publishing; 2021.
56. Tranquada RE. Lactic acidosis. *Calif Med*. 1964;101(6):450.
57. Zhao J, Xu L, Wang Y, Zhao X, Wang J, Garza E, et al. Homofermentative production of optically pure L-lactic acid from xylose by genetically engineered *Escherichia coli* B. *Microb Cell Fact*. 2013;12(1):1–6.
58. Garlotta D. A literature review of poly (lactic acid). *J Polym Environ*. 2001;9(2):63–84.
59. Kaplan DL. Introduction to biopolymers from renewable resources. *Biopolymers from renewable resources*. Springer; 1998. pp. 1–29.
60. Mehta R, Kumar V, Bhunia H, Upadhyay S. Synthesis of poly (lactic acid): a review. *J Macromolecular Sci Part C: Polym Reviews*. 2005;45(4):325–49.
61. Jiang X, Luo Y, Tian X, Huang D, Reddy N, Yang Y. Chemical structure of poly (lactic acid). *Poly (Lactic Acid) Synthesis, Structures, Properties, Processing, and Applications*. 2010:67–82.
62. Fambri L, Migliaresi C. Crystallization and thermal properties. *Poly (Lactic Acid) Synthesis, Structures, Properties, Processing, Applications, and End of Life*. 2022:135 – 51.
63. Vaid R, Yildirim E, Pasquini MA, King MW. Hydrolytic degradation of polylactic acid fibers as a function of pH and exposure time. *Molecules*. 2021;26(24):7554.
64. Gonçalves CM, Coutinho JoA, Marrucho IM. Optical properties. *Poly (Lactic Acid) Synthesis, Structures, Properties, Processing, and Applications*. 2010:97–112.
65. Perego G, Cella GD. Mechanical properties. *Poly (Lactic Acid) Synthesis, Structures, Properties, Processing, and Applications*. 2010:141 – 53.
66. Almenar E, Auras R. Permeation, sorption, and diffusion in poly (lactic acid). *Poly (lactic acid) synthesis, structure, properties, processing and applications* Wiley, Hoboken. 2010:155 – 79.
67. Dorgan JR. Rheology of poly (lactic acid). *Poly (Lactic Acid) Synthesis, Structures, Properties, Processing, Applications, and End of Life*. 2022:153 – 67.
68. DeStefano V, Khan S, Tabada A. Applications of PLA in modern medicine. *Eng Regeneration*. 2020;1:76–87.
69. Feng P, Jia J, Liu M, Peng S, Zhao Z, Shuai C. Degradation mechanisms and acceleration strategies of poly (lactic acid) scaffold for bone regeneration. *Mater Design*. 2021;210:110066. <https://doi.org/10.1016/j.matdes.2021.110066>.
70. Alexis F. Factors affecting the degradation and drug-release mechanism of poly(lactic acid) and poly[(lactic acid)-co-(glycolic acid)]. *Polym Int*. 2005;54(1):36–46. <https://doi.org/10.1002/pi.1697>.
71. Zaaba NF, Jaafar M. A review on degradation mechanisms of polylactic acid: Hydrolytic, photodegradative, microbial, and enzymatic degradation. *Polym Eng Sci*. 2020;60(9):2061–75. <https://doi.org/10.1002/pen.25511>.
72. Green JJ, Elisseeff JH. Mimicking biological functionality with polymers for biomedical applications. *Nature*. 2016;540(7633):386–94.
73. Freeman R, Boekhoven J, Dickerson MB, Naik RR, Stupp SI. Biopolymers and supramolecular polymers as biomaterials for biomedical applications. *MRS Bull*. 2015;40(12):1089–101.
74. Chen F-M, Liu X. Advancing biomaterials of human origin for tissue engineering. *Prog Polym Sci*. 2016;53:86–168.
75. Heidari BS, Ruan R, Vahabi E, Chen P, De-Juan-Pardo EM, Zheng M, et al. Natural, synthetic and commercially-available biopolymers used to regenerate tendons and ligaments. *Bioactive Mater*. 2023;19:179–97.
76. Gardikiotis I, Cojocar F-D, Mihai C-T, Balan V, Dodi G. Borrowing the features of biopolymers for emerging Wound Healing Dressings: a review. *Int J Mol Sci*. 2022;23(15):8778.
77. Haugen HJ, Lyngstadaas SP, Rossi F, Perale G. Bone grafts: which is the ideal biomaterial? *J Clin Periodontol*. 2019;46:92–102.
78. Klar AS, Zimoch J, Biedermann T. Skin tissue engineering: application of adipose-derived stem cells. *BioMed research international*. 2017;2017.
79. Saltzman WM, Kyriakides TR. Chapter 16 - Cell interactions with polymers. In: Lanza R, Langer R, Vacanti JP, Atala A, editors. *Principles of Tissue Engineering (Fifth Edition)*. Academic Press; 2020. p. 275 – 93.
80. Surendren A, Cheekuramelli NS, Magisetty RP. 22 - biodegradable polymer blends for tissue engineering. In: Mavinkere Rangappa S, Parameswaranpillai J, Siengchin S, Ramesh M, editors. *Biodegradable polymers, blends and composites*. Woodhead Publishing; 2022. pp. 591–609.
81. Ramot Y, Haim-Zada M, Domb AJ, Nyska A. Biocompatibility and safety of PLA and its copolymers. *Adv Drug Deliv Rev*. 2016;107. <https://doi.org/10.1016/j.addr.2016.03.012>. :153 – 62.
82. Berthiaume F, Maguire TJ, Yarmush ML. Tissue engineering and regenerative medicine: history, progress, and challenges. *Annual Rev Chem Biomol Eng*. 2011;2:403–30.
83. Pearson R, Bhandari R, Quirk R, Shakesheff KM. Recent advances in tissue engineering. *J Long Term Eff Med Implants*. 2017;27:2–4.
84. Auras RA, Lim L-T, Selke SE, Tsuji H. Poly (lactic acid): synthesis, structures, properties, processing, and applications. *John Wiley & Sons*; 2011.
85. Zhang WJ, Liu W, Cui L, Cao Y. Tissue engineering of blood vessel. *J Cell Mol Med*. 2007;11(5):945–57.
86. Ding H, Cheng Y, Niu X, Hu Y. Application of electrospun nanofibers in bone, cartilage and osteochondral tissue engineering. *J Biomater Sci Polym Ed*. 2020;32(4):536–61.
87. Alonzo M, Anilkumar S, Roman B, Tasnim N, Joddar B. 3D bioprinting of cardiac tissue and cardiac stem cell therapy. *Translational Res*. 2019;211:64–83.
88. Wu Y, Zhang X, Zhao Q, Tan B, Chen X, Liao J. Role of hydrogels in bone tissue engineering: how properties shape regeneration. *J Biomed Nanotechnol*. 2020;16(12):1667–86.
89. Dehqan Niri A, Karimi Zarchi AA, Ghadiri Harati P, Salimi A, Mujokoro B. Tissue engineering scaffolds in the treatment of brain disorders in geriatric patients. *Artif Organs*. 2019;43(10):947–60.
90. Adamowicz J, Kuffel B, Van Breda SV, Pokrwczynska M, Drewa T. Reconstructive urology and tissue engineering: converging developmental paths. *J Tissue Eng Regen Med*. 2019;13(3):522–33.
91. Koch KL, Bitar KN, Fortunato JE. Tissue engineering for neuromuscular disorders of the gastrointestinal tract. *World J Gastroenterology: WJG*. 2012;18(47):6918.
92. Langer R, Vacanti J. Advances in tissue engineering. *J Pediatr Surg*. 2016;51(1):8–12.
93. Cheng Y, Deng S, Chen P, Ruan R. Poly(lactic acid) (PLA) synthesis and modifications: a review. *Front Chem China*. 2009;4(3):259–64.
94. Saini P, Arora M, Kumar MR. Poly (lactic acid) blends in biomedical applications. *Adv Drug Deliv Rev*. 2016;107:47–59.
95. Yu L, Petinakis E, Dean K, Liu H, Yuan Q. Enhancing compatibilizer function by controlled distribution in hydrophobic polylactic acid/hydrophilic starch blends. *J Appl Polym Sci*. 2011;119(4):2189–95.
96. Muir VG, Burdick JA. Chemically modified biopolymers for the formation of biomedical hydrogels. *Chem Rev*. 2020;121(18):10908–49.
97. Yoshikawa H, Myoui A. Bone tissue engineering with porous hydroxyapatite ceramics. *J Artif Organs*. 2005;8(3):131–6.
98. Mondal S, Nguyen TP, Hoang G, Manivasagan P, Kim MH, Nam SY, et al. Hydroxyapatite nano bioceramics optimized 3D printed poly lactic acid scaffold for bone tissue engineering application. *Ceram Int*. 2020;46(3):3443–55.
99. Flores-Sánchez MG, Islas - Arteaga NC, Raya - Rivera AM, Esquiliano - Rendón DR, Morales - Corona J, Uribe - Juárez OE, et al. Effect of a plasma synthesized polypyrrole coverage on polylactic acid/hydroxyapatite scaffolds for bone tissue engineering. *J Biomedical Mater Res Part A*. 2021;109(11):2199–211.
100. Ma H, Su W, Tai Z, Sun D, Yan X, Liu B, et al. Preparation and cytocompatibility of polylactic acid/hydroxyapatite/graphene oxide nanocomposite fibrous membrane. *Chin Sci Bull*. 2012;57(23):3051–8.
101. Rong Z, Zeng W, Kuang Y, Zhang J, Liu X, Lu Y, et al. Enhanced bioactivity of osteoblast-like cells on poly (lactic acid)/poly (methyl methacrylate)/

- nano-hydroxyapatite scaffolds for bone tissue engineering. *Fibers Polym.* 2015;16(2):245–53.
102. Abdal-hay A, Sheikh FA, Lim JK. Air jet spinning of hydroxyapatite/poly (lactic acid) hybrid nanocomposite membrane mats for bone tissue engineering. *Colloids Surf B.* 2013;102:635–43.
103. Yoshikawa H, Tamai N, Murase T, Myoui A. Interconnected porous hydroxyapatite ceramics for bone tissue engineering. *J Royal Soc Interface.* 2009;6(suppl3):341–58.
104. Shi H, Zhou Z, Li W, Fan Y, Li Z, Wei J. Hydroxyapatite based materials for bone tissue engineering: a brief and comprehensive introduction. *Crystals.* 2021;11(2):149.
105. Bahú JO, Melo de Andrade LR, Crivellin S, Khouri NG, Sousa SO, Fernandes LMI, et al. Rotary jet spinning (RJS): a key process to produce Biopolymeric Wound Dressings. *Pharmaceutics.* 2022;14(11). <https://doi.org/10.3390/pharmaceutics14112500>.
106. Rosa JC, Bonvent JJ, Santos AR. Jr. Poly (ϵ -caprolactone)/Poly (lactic acid) fibers produced by rotary jet spinning for skin dressing with antimicrobial activity. *J Biomater Appl.* 2022;36(9):1641–51. <https://doi.org/10.1177/08853282211064946>.
107. Barbosa KA, Rodrigues ICP, Tamborlin L, Luchessi AD, Lopes ÉSN, Gabriel LP. Rotary jet-spun curcumin-loaded poly L-lactic acid membranes for wound-healing applications. *J Mater Res Technol.* 2022;18:3273–82. <https://doi.org/10.1016/j.jmrt.2022.03.136>.
108. Rodchanasuriporn W, Seadan M, Suttiruengwong S. Properties of non-woven polylactic acid fibers prepared by the rotational jet spinning method. *Mater Today Sustain.* 2020;10:100046. <https://doi.org/10.1016/j.mtsust.2020.100046>.
109. Ravishankar P, Khang A, Laredo M, Balachandran K. Using Dimensionless numbers to Predict Centrifugal Jet-Spun Nanofiber morphology. *J Nanomaterials.* 2019;2019:4639658. <https://doi.org/10.1155/2019/4639658>.
110. Xia L, Lu L, Liang Y, Cheng B. Fabrication of centrifugally spun prepared poly(lactic acid)/gelatin/ciprofloxacin nanofibers for antimicrobial wound dressing. *RSC Adv.* 2019;9(61):35328–35. <https://doi.org/10.1039/c9ra07826f>.
111. Corcione CE, Gervaso F, Scalera F, Padmanabhan SK, Madaghiele M, Montagna F, et al. Highly loaded hydroxyapatite microsphere/PLA porous scaffolds obtained by fused deposition modelling. *Ceram Int.* 2019;45(2):2803–10.
112. Mohandesnezhad S, Pilehvar-Soltanahmadi Y, Alizadeh E, Goodarzi A, Davaran S, Khatamian M, et al. In vitro evaluation of Zeolite-nHA blended PCL/PLA nanofibers for dental tissue engineering. *Mater Chem Phys.* 2020;252:123152.
113. Tayton E, Purcell M, Aarvold A, Smith J, Briscoe A, Kanczler J, et al. A comparison of polymer and polymer–hydroxyapatite composite tissue engineered scaffolds for use in bone regeneration. An in vitro and in vivo study. *J Biomedical Mater Res Part A.* 2014;102(8):2613–24.
114. Tamai N, Myoui A, Hirao M, Kaito T, Ochi T, Tanaka J, et al. A new biotechnology for articular cartilage repair: subchondral implantation of a composite of interconnected porous hydroxyapatite, synthetic polymer (PLA-PEG), and bone morphogenetic protein-2 (rhBMP-2). *Osteoarthritis Cartil.* 2005;13(5):405–17.
115. Moran JM, Pazzano D, Bonassar LJ. Characterization of polylactic acid–polyglycolic acid composites for cartilage tissue engineering. *Tissue Eng.* 2003;9(1):63–70.
116. Tuli R, Li W-J, Tuan RS. Current state of cartilage tissue engineering. *Arthritis Res Ther.* 2003;5(5):1–4.
117. Luo X, Zhou G, Liu W, Zhang WJ, Cen L, Cui L, et al. In vitro precultivation alleviates post-implantation inflammation and enhances development of tissue-engineered tubular cartilage. *Biomed Mater.* 2009;4(2):025006.
118. Lavik E, Teng YD, Snyder E, Langer R. Seeding neural stem cells on scaffolds of PGA, PLA, and their copolymers. *Neural stem cells: methods and protocols.* Springer; 2002. pp. 89–96.
119. Xu H, Han D, Dong JS, Shen GX, Chai G, Yu ZY, et al. Rapid prototyped PGA/PLA scaffolds in the reconstruction of mandibular condyle bone defects. *Int J Med Rob Comput Assist Surg.* 2010;6(1):66–72.
120. Agrawal CM, Ray RB. Biodegradable polymeric scaffolds for musculoskeletal tissue engineering. *Journal of Biomedical Materials Research: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials.* 2001;55(2):141–50.
121. Giraud M-N, Armbruster C, Carrel T, Tevaearai HT. Current state of the art in myocardial tissue engineering. *Tissue Eng.* 2007;13(8):1825–36.
122. Reichardt A, Arshi A, Schuster P, Polchow B, Shakibaei M, Gries T, et al. Custom-made generation of three-dimensional nonwovens composed of polyglycolide or polylactide for the cardiovascular tissue engineering. *J Biomaterials Tissue Eng.* 2012;2(4):322–9.
123. Cantón I, Mckean R, Charnley M, Blackwood KA, Fiorica C, Ryan AJ, et al. Development of an ibuprofen-releasing biodegradable PLA/PGA electrospun scaffold for tissue regeneration. *Biotechnol Bioeng.* 2010;105(2):396–408.
124. Eğri S, Eczacıoğlu N. Sequential VEGF. BMP-2 releasing PLA-PEG-PLA scaffolds for bone tissue engineering: I. Design and in vitro tests. *Artif cells Nanomed Biotechnol.* 2017;45(2):321–9.
125. Kang Y, Chen P, Shi X, Zhang G, Wang C. Multilevel structural stereocomplex polylactic acid/collagen membranes by pattern electrospinning for tissue engineering. *Polymer.* 2018;156:250–60.
126. Khatsee S, Daranarong D, Punyodom W, Worajittiphon P. Electrospinning polymer blend of PLA and PBAT: electrospinnability–solubility map and effect of polymer solution parameters toward application as antibiotic-carrier mats. *J Appl Polym Sci.* 2018;135(28):46486.
127. Hongdilokkul P, Keeratipinit K, Chawthai S, Hararak B, Seadan M, Suttiruengwong S. A study on properties of PLA/PBAT from blown film process. *IOP conference series: materials science and engineering: IOP Publishing; 2015.* p. 012112.
128. Jadoun S. POLYLACTIDE (PLA) BASED NANOCOMPOSITES FOR APPLICATIONS IN ANTIBACTERIAL/MICROBIAL AND BIOMEDICAL ENGINEERING. *BIOCOMPOSITES IN BIO-MEDICINE.*69.
129. Yan D, Wang Z, Guo Z, Ma Y, Wang C, Tan H, et al. Study on the properties of PLA/PBAT composite modified by nanohydroxyapatite. *J Mater Res Technol.* 2020;9(5):11895–904. <https://doi.org/10.1016/j.jmrt.2020.08.062>.
130. Kang Y, Chen P, Shi X, Zhang G, Wang C. Preparation of open-porous stereocomplex PLA/PBAT scaffolds and correlation between their morphology, mechanical behavior, and cell compatibility. *RSC Adv.* 2018;8(23):12933–43.
131. Hernández-López M, Correa-Pacheco ZN, Bautista-Baños S, Zavaleta-Avejar L, Benítez-Jiménez JJ, Sabino-Gutiérrez MA, et al. Bio-based composite fibers from pine essential oil and PLA/PBAT polymer blend. Morphological, physicochemical, thermal and mechanical characterization. *Mater Chem Phys.* 2019;234:345–53.
132. Nofar M, Tabatabaei A, Sojoudiasli H, Park C, Carreau P, Heuzey M-C, et al. Mechanical and bead foaming behavior of PLA-PBAT and PLA-PBSA blends with different morphologies. *Eur Polymer J.* 2017;90:231–44.
133. Cui H, Shao J, Wang Y, Zhang P, Chen X, Wei Y. PLA-PEG-PLA and its electroactive tetraaniline copolymer as multi-interactive injectable hydrogels for tissue engineering. *Biomacromolecules.* 2013;14(6):1904–12.
134. Atiqah MN, Sharifah I, Yose F, Maizatunisa O, Norhashimah S. Characterization of poly (lactic acid)/poly (ethylene) glycol blends prepared for melt drawn spinning process. *Materials Today: Proceedings.* 2019;17:889 – 97.
135. Serra T, Ortiz-Hernandez M, Engel E, Planell JA, Navarro M. Relevance of PEG in PLA-based blends for tissue engineering 3D-printed scaffolds. *Mater Sci Engineering: C.* 2014;38:55–62.
136. Tessmar JK, Göpferich AM. Customized PEG-derived copolymers for tissue - engineering applications. *Macromol Biosci.* 2007;7(1):23–39.
137. Huang R, Zhu X, Zhao T, Wan A. Preparation of tissue engineering porous scaffold with poly (lactic acid) and polyethylene glycol solution blend by solvent-casting/particulate-leaching. *Mater Res Express.* 2014;1(4):045403.
138. Zhao X, Hadjiaryrou M. Induction of cell migration in vitro by an electrospun PDGF-BB/PLGA/PEG-PLA nanofibrous scaffold. *J Biomed Nanotechnol.* 2011;7(6):823–9.
139. Zhong T, Deng C, Gao Y, Chen M, Zuo B. Studies of in situ-forming hydrogels by blending PLA - PEG - PLA copolymer with silk fibroin solution. *J biomedical Mater Res Part A.* 2012;100(8):1983–9.
140. Bhaskar B, Owen R, Bahmaee H, Wally Z, Sreenivasa Rao P, Reilly GC. Composite porous scaffold of PEG/PLA support improved bone matrix deposition in vitro compared to PLA-only scaffolds. *J Biomedical Mater Res Part A.* 2018;106(5):1334–40.
141. Ni P, Fu S, Fan M, Guo G, Shi S, Peng J, et al. Preparation of poly (ethylene glycol)/polylactide hybrid fibrous scaffolds for bone tissue engineering. *Int J Nanomed.* 2011;6:3065.
142. Rahman CV, Kuhn G, White LJ, Kirby GT, Varghese OP, McLaren JS, et al. PLGA/PEG-hydrogel composite scaffolds with controllable mechanical properties. *J Biomedical Mater Res Part B: Appl Biomaterials.* 2013;101(4):648–55.
143. Sugiarto S, Leow Y, Tan CL, Wang G, Kai D. How far is lignin from being a biomedical material? *Bioactive Mater.* 2022;8:71–94. <https://doi.org/10.1016/j.bioactmat.2021.06.023>.
144. Byrne CE, Astete CE, Vaithyanathan M, Melvin AT, Moradipour M, Rankin SE, et al. Lignin-graft-PLGA drug-delivery system improves efficacy of

- MEK1/2 inhibitors in triple-negative breast cancer cell line. *Nanomedicine*. 2020;15(10):981–1000.
145. Luzi F, Tortorella I, Di Michele A, Dominici F, Argentati C, Morena F, et al. Novel nanocomposite PLA films with lignin/zinc oxide hybrids: design, characterization, interaction with mesenchymal stem cells. *Nanomaterials*. 2020;10(11):2176.
146. Boni R, Ali A, Shavandi A, Clarkson AN. Current and novel polymeric biomaterials for neural tissue engineering. *J Biomed Sci*. 2018;25(1):1–21.
147. Liang R, Yang X, Yew PYM, Sugiarto S, Zhu Q, Zhao J et al. PLA-lignin nanofibers as antioxidant biomaterials for cartilage regeneration and osteoarthritis treatment. 2022.
148. Nan N, Hu W, Wang J. Lignin-based porous biomaterials for Medical and Pharmaceutical Applications. *Biomedicines*. 2022;10(4):747.
149. Zhang N. Preparation of nanofibre material based on electrospinning technology and its application in rehabilitation of lower limb joint motion. *Int J Nanotechnol*. 2020;17(2–6):325–38.
150. Liang Y, Goh JC-H. Polypyrrole-incorporated conducting constructs for tissue engineering applications: a review. *Bioelectricity*. 2020;2(2):101–19.
151. Tian L, Prabhakaran MP, Hu J, Chen M, Besenbacher F, Ramakrishna S. Synergistic effect of topography, surface chemistry and conductivity of the electrospun nanofibrous scaffold on cellular response of PC12 cells. *Colloids Surf B*. 2016;145:420–9.
152. Shu B, Liu XB, Zhou JF, Huang H, Wang JY, Sun XD, et al. Polypyrrole/poly(lactic acid) nanofibrous scaffold cotransplanted with bone marrow stromal cells promotes the functional recovery of spinal cord injury in rats. *CNS Neurosci Ther*. 2019;25(9):951–64.
153. Zhou X, Yang A, Huang Z, Yin G, Pu X, Jin J. Enhancement of neurite adhesion, alignment and elongation on conductive polypyrrole-poly (lactide acid) fibers with cell-derived extracellular matrix. *Colloids Surf B*. 2017;149:217–25.
154. Zhou J-f, Wang Y-g, Cheng L, Wu Z, Sun X-d, Peng J. Preparation of polypyrrole-embedded electrospun poly (lactic acid) nanofibrous scaffolds for nerve tissue engineering. *Neural regeneration research*. 2016;11(10):1644.
155. Chen G, Ushida T, Tateishi T. Hybrid biomaterials for tissue engineering: a preparative method for PLA or PLGA–collagen hybrid sponges. *Adv Mater*. 2000;12(6):455–7.
156. Sivashankari P, Prabaharan M. Prospects of chitosan-based scaffolds for growth factor release in tissue engineering. *Int J Biol Macromol*. 2016;93:1382–9.
157. Shalumon K, Sathish D, Nair S, Chennazhi K, Tamura H, Jayakumar R. Fabrication of aligned poly (lactic acid)-chitosan nanofibers by novel parallel blade collector method for skin tissue engineering. *J Biomed Nanotechnol*. 2012;8(3):405–16.
158. Kim I-Y, Seo S-J, Moon H-S, Yoo M-K, Park I-Y, Kim B-C, et al. Chitosan and its derivatives for tissue engineering applications. *Biotechnol Adv*. 2008;26(1):1–21.
159. Radwan-Pragłowska J, Janus Ł, Piątkowski M, Bogdał D, Matysek D. Hybrid bilayer PLA/chitosan nanofibrous scaffolds doped with ZnO, Fe₃O₄, and Au nanoparticles with bioactive properties for skin tissue engineering. *Polymers*. 2020;12(1):159.
160. Zeng S, Ye J, Cui Z, Si J, Wang Q, Wang X, et al. Surface biofunctionalization of three-dimensional porous poly (lactic acid) scaffold using chitosan/OGP coating for bone tissue engineering. *Mater Sci Engineering: C*. 2017;77:92–101.
161. Hu X, He J, Yong X, Lu J, Xiao J, Liao Y, et al. Biodegradable poly (lactic acid-co-trimethylene carbonate)/chitosan microsphere scaffold with shape-memory effect for bone tissue engineering. *Colloids Surf B*. 2020;195:11218.
162. Xu J, Zhang J, Gao W, Liang H, Wang H, Li J. Preparation of chitosan/PLA blend micro/nanofibers by electrospinning. *Mater Lett*. 2009;63(8):658–60. <https://doi.org/10.1016/j.matlet.2008.12.014>.
163. Shen R, Xu W, Xue Y, Chen L, Ye H, Zhong E, et al. The use of chitosan/PLA nano-fibers by emulsion electrospinning for periodontal tissue engineering. *Artif cells Nanomed Biotechnol*. 2018;46(sup2):419–30.
164. Radwan-Pragłowska J, Janus Ł, Piątkowski M, Bogdał D, Matysek D. 3D hierarchical, nanostructured chitosan/PLA/HA scaffolds doped with TiO₂/Au/Pt NPs with tunable properties for guided bone tissue engineering. *Polymers*. 2020;12(4):792.
165. Liu Y, Wang S, Zhang R. Composite poly (lactic acid)/chitosan nanofibrous scaffolds for cardiac tissue engineering. *Int J Biol Macromol*. 2017;103:1130–7.
166. Hassanajili S, Karami-Pour A, Oryan A, Talaei-Khozani T. Preparation and characterization of PLA/PCL/HA composite scaffolds using indirect 3D printing for bone tissue engineering. *Mater Sci Engineering: C*. 2019;104:109960.
167. Arif ZU, Khalid MY, Noroozi R, Sadeghianmarjan A, Jalalvand M, Hossain M. Recent advances in 3D-printed polylactide and polycaprolactone-based biomaterials for tissue engineering applications. *International Journal of Biological Macromolecules*. 2022.
168. Sartore L, Inverardi N, Pandini S, Bignotti F, Chiellini F. PLA/PCL-based foams as scaffolds for tissue engineering applications. *Materials Today: Proceedings*. 2019;7:410–7. doi: <https://doi.org/10.1016/j.matpr.2018.11.103>.
169. Patrício T, Domingos M, Gloria A, D'Amora U, Coelho J, Bártolo P. Fabrication and characterisation of PCL and PCL/PLA scaffolds for tissue engineering. *Rapid Prototyp J*. 2014;20(2):145–56.
170. Vaz C, Van Tuijl S, Bouten C, Baaijens F. Design of scaffolds for blood vessel tissue engineering using a multi-layering electrospinning technique. *Acta Biomater*. 2005;1(5):575–82.
171. Sankaran KK, Vasanthan KS, Krishnan UM, Sethuraman S. Development and evaluation of axially aligned nanofibres for blood vessel tissue engineering. *J Tissue Eng Regen Med*. 2014;8(8):640–51.
172. Buscemi S, Palumbo V, Maffongelli A, Fazzotta S, Palumbo F, Licciardi M et al. Electrospun PHEA-PLA/PCL scaffold for vascular regeneration: a preliminary in vivo evaluation. *Transplantation Proceedings*; Elsevier; 2017. p. 716 – 21.
173. Pitarresi G, Fiorica C, Palumbo FS, Rigogliuso S, Ghersi G, Giammona G. Heparin functionalized polyaspartamide/polyester scaffold for potential blood vessel regeneration. *J Biomedical Mater Res Part A*. 2014;102(5):1334–41.
174. Li C, Wang F, Douglas G, Zhang Z, Guidoin R, Wang L. Comprehensive mechanical characterization of PLA fabric combined with PCL to form a composite structure vascular graft. *J Mech Behav Biomed Mater*. 2017;69:39–49.
175. Radulescu D, Dhar S, Young CM, Taylor DW, Trost H-J, Hayes DJ, et al. Tissue engineering scaffolds for nerve regeneration manufactured by ink-jet technology. *Mater Sci Engineering: C*. 2007;27(3):534–9.
176. Mondal S, Nguyen TP, Pham VH, Hoang G, Manivasagan P, Kim MH, et al. Hydroxyapatite nano bioceramics optimized 3D printed poly lactic acid scaffold for bone tissue engineering application. *Ceram Int*. 2020;46(3):3443–55. <https://doi.org/10.1016/j.ceramint.2019.10.057>.
177. Rong Z, Zeng W, Kuang Y, Zhang J, Liu X, Lu Y, et al. Enhanced bioactivity of osteoblast-like cells on poly(lactic acid)/poly(methyl methacrylate)/nano-hydroxyapatite scaffolds for bone tissue engineering. *Fibers Polym*. 2015;16(2):245–53. <https://doi.org/10.1007/s12221-015-0245-0>.
178. Abdal-hay A, Sheikh FA, Lim JK. Air jet spinning of hydroxyapatite/poly(lactic acid) hybrid nanocomposite membrane mats for bone tissue engineering. *Colloids Surf B*. 2013;102:635–43. <https://doi.org/10.1016/j.colsurfb.2012.09.017>.
179. Yoshikawa H, Tamai N, Murase T, Myoui A. Interconnected porous hydroxyapatite ceramics for bone tissue engineering. *J R Soc Interface*. 2009;6(Suppl 3):341–8. <https://doi.org/10.1098/rsif.2008.0425.focus>.
180. Tayton E, Purcell M, Aarvold A, Smith JO, Briscoe A, Kanczler JM, et al. A comparison of polymer and polymer-hydroxyapatite composite tissue engineered scaffolds for use in bone regeneration. An in vitro and in vivo study. *J Biomed Mater Res A*. 2014;102(8):2613–24. <https://doi.org/10.1002/jbma.34926>.
181. Tamai N, Myoui A, Hirao M, Kaito T, Ochi T, Tanaka J, et al. A new biotechnology for articular cartilage repair: subchondral implantation of a composite of interconnected porous hydroxyapatite, synthetic polymer (PLA-PEG), and bone morphogenetic protein-2 (rhBMP-2). *Osteoarthritis Cartilage*. 2005;13(5):405–17. <https://doi.org/10.1016/j.joca.2004.12.014>.
182. Moran JM, Pazzano D, Bonassar LJ. Characterization of poly(lactic acid)-polyglycolic acid composites for cartilage tissue engineering. *Tissue Eng*. 2003;9(1):63–70. <https://doi.org/10.1089/107632703762687546>.
183. Tuli R, Li WJ, Tuan RS. Current state of cartilage tissue engineering. *Arthritis Res Ther*. 2003;5(5):235–8. <https://doi.org/10.1186/ar991>.
184. Agrawal CM, Ray RB. Biodegradable polymeric scaffolds for musculoskeletal tissue engineering. *J Biomed Mater Res*. 2001;55(2):141–50. [https://doi.org/10.1002/1097-4636\(200105\)55:2<141::aid-jbmm1000>3.0.co;2-j](https://doi.org/10.1002/1097-4636(200105)55:2<141::aid-jbmm1000>3.0.co;2-j).
185. Cantón I, McKean R, Charnley M, Blackwood KA, Fiorica C, Ryan AJ, et al. Development of an ibuprofen-releasing biodegradable PLA/PGA electrospun scaffold for tissue regeneration. *Biotechnol Bioeng*. 2010;105(2):396–408. <https://doi.org/10.1002/bit.22530>.
186. Khatsee S, Daranarong D, Punyodom W, Worajittiphon P. Electrospinning polymer blend of PLA and PBAT: electrospinnability–solubility map and effect of polymer solution parameters toward application as antibiotic-carrier mats. *J Appl Polym Sci*. 2018;135(28):46486. <https://doi.org/10.1002/app.46486>.
187. Nofar M, Tabatabaei A, Sojoudiasli H, Park CB, Carreau PJ, Heuzey MC, et al. Mechanical and bead foaming behavior of PLA-PBAT and PLA-PBSA blends

- with different morphologies. *Eur Polymer J.* 2017;90:231–44. <https://doi.org/10.1016/j.eurpolymj.2017.03.031>.
188. Ni P, Fu S, Fan M, Guo G, Shi S, Peng J, et al. Preparation of poly(ethylene glycol)/polylactide hybrid fibrous scaffolds for bone tissue engineering. *Int J Nanomedicine.* 2011;6:3065–75. <https://doi.org/10.2147/ijn.S25297>.
189. Bhaskar B, Owen R, Bahmaee H, Wally Z, Sreenivasa Rao P, Reilly GC. Composite porous scaffold of PEG/PLA support improved bone matrix deposition in vitro compared to PLA-only scaffolds. *J Biomed Mater Res A.* 2018;106(5):1334–40. <https://doi.org/10.1002/jbma.36336>.
190. Tessmar JK, Göpferich AM. Customized PEG-Derived copolymers for Tissue-Engineering Applications. *Macromol Biosci.* 2007;7(1):23–39. <https://doi.org/10.1002/mabi.200600096>.
191. Serra T, Ortiz-Hernandez M, Engel E, Planell JA, Navarro M. Relevance of PEG in PLA-based blends for tissue engineering 3D-printed scaffolds. *Mater Sci Eng C Mater Biol Appl.* 2014;38:55–62. <https://doi.org/10.1016/j.msec.2014.01.003>.
192. Ngadimin KD, Stokes A, Gentile P, Ferreira AM. Biomimetic hydrogels designed for cartilage tissue engineering. *Biomaterials Sci.* 2021;9(12):4246–59. <https://doi.org/10.1039/D0BM01852J>.
193. Liang R, Yang X, Yew PYM, Sugianto S, Zhu Q, Zhao J, et al. PLA-lignin nanofibers as antioxidant biomaterials for cartilage regeneration and osteoarthritis treatment. *J Nanobiotechnol.* 2022;20(1):327. <https://doi.org/10.1186/s12951-022-01534-2>.
194. Boni R, Ali A, Shavandi A, Clarkson AN. Current and novel polymeric biomaterials for neural tissue engineering. *J Biomed Sci.* 2018;25(11):90. <https://doi.org/10.1186/s12929-018-0491-8>.
195. Luzi F, Tortorella I, Di Michele A, Dominici F, Argentati C, Morena F, et al. Novel nanocomposite PLA Films with Lignin/Zinc oxide hybrids: design, characterization, Interaction with mesenchymal stem cells. *Nanomaterials.* 2020. <https://doi.org/10.3390/nano10112176>.
196. Liu R, Dai L, Xu C, Wang K, Zheng C, Si C. Lignin-based Micro- and nanomaterials and their composites in Biomedical Applications. *Chemschem.* 2020;13(17):4266–83. <https://doi.org/10.1002/cssc.202000783>.
197. Raynald, Shu B, Liu XB, Zhou JF, Huang H, Wang JY, et al. Polypyrrole/polylactic acid nanofibrous scaffold cotransplanted with bone marrow stromal cells promotes the functional recovery of spinal cord injury in rats. *CNS Neurosci Ther.* 2019;25(9):951–64. <https://doi.org/10.1111/cns.13135>.
198. Liang Y, Goh JC. Polypyrrole-Incorporated conducting constructs for tissue Engineering Applications: a review. *Bioelectricity.* 2020;2(2):101–19. <https://doi.org/10.1089/bioe.2020.0010>.
199. Liu Y, Wang S, Zhang R. Composite poly(lactic acid)/chitosan nanofibrous scaffolds for cardiac tissue engineering. *Int J Biol Macromol.* 2017;103:1130–7. <https://doi.org/10.1016/j.jbiomac.2017.05.101>.
200. Sivashankari PR, Prabakaran M. Prospects of chitosan-based scaffolds for growth factor release in tissue engineering. *Int J Biol Macromol.* 2016;93(Pt B):1382–9. doi: <https://doi.org/10.1016/j.jbiomac.2016.02.043>.
201. Zeng S, Ye J, Cui Z, Si J, Wang Q, Wang X, et al. Surface biofunctionalization of three-dimensional porous poly(lactic acid) scaffold using chitosan/OGP coating for bone tissue engineering. *Mater Sci Eng C Mater Biol Appl.* 2017;77:92–101. <https://doi.org/10.1016/j.msec.2017.03.220>.
202. Li C, Wang F, Douglas G, Zhang Z, Guidoin R, Wang L. Comprehensive mechanical characterization of PLA fabric combined with PCL to form a composite structure vascular graft. *J Mech Behav Biomed Mater.* 2017;69:39–49. <https://doi.org/10.1016/j.jmbm.2016.11.005>.
203. Vaz CM, van Tuijl S, Bouten CV, Baaijens FP. Design of scaffolds for blood vessel tissue engineering using a multi-layering electrospinning technique. *Acta Biomater.* 2005;1(5):575–82. <https://doi.org/10.1016/j.actbio.2005.06.006>.
204. Hassanajili S, Karami-Pour A, Oryan A, Talaei-Khozani T. Preparation and characterization of PLA/PCL/HA composite scaffolds using indirect 3D printing for bone tissue engineering. *Mater Sci Eng C Mater Biol Appl.* 2019;104:109960. <https://doi.org/10.1016/j.msec.2019.109960>.
205. Ren Q, Li W, Cui S, Ma W, Zhu X, Wu M, et al. Improved thermal insulation and compressive property of bimodal poly (lactic acid)/cellulose nanocomposite foams. *Carbohydr Polym.* 2023;302:120419. <https://doi.org/10.1016/j.carbpol.2022.120419>.
206. Gunes OC, Kara A, Baysan G, Bugra Husemoglu R, Akokay P, Ziyilan Albayrak A, et al. Fabrication of 3D printed poly(lactic acid) strut and wet-electrospun cellulose nano fiber reinforced chitosan-collagen hydrogel composite scaffolds for meniscus tissue engineering. *J Biomater Appl.* 2022;37(4):683–97. <https://doi.org/10.1177/08853282221109339>.
207. Jafari A, Mirzaei H, Shafei MA, Fakhri V, Yazdanbakhsh A, Pirouzar V, et al. Conductive poly(ϵ -caprolactone)/polylactic acid scaffolds for tissue engineering applications: synergy effect of zirconium nanoparticles and polypyrrole. *Polym Adv Technol.* 2022;33(5):1427–41. <https://doi.org/10.1002/pat.5611>.
208. Ye G, Li Z, Chen B, Bai X, Chen X, Hu Y. Performance of polylactic acid/polycaprolactone/microcrystalline cellulose biocomposites with different filler contents and maleic anhydride compatibilization. *Polym Compos.* 2022;43(8):5179–88. <https://doi.org/10.1002/pc.26807>.
209. Mehrpouya M, Vahabi H, Janbaz S, Darafsheh A, Mazur TR, Ramakrishna S. 4D printing of shape memory polylactic acid (PLA). *Polymer.* 2021;230:124080. <https://doi.org/10.1016/j.polymer.2021.124080>.
210. Lin C, Liu L, Liu Y, Leng J. 4D printing of shape memory polybutylene succinate/poly(lactic acid) (PBS/PLA) and its potential applications. *Compos Struct.* 2022;279:114729. <https://doi.org/10.1016/j.compstruct.2021.114729>.
211. Nonkrathok W, Trongsatitkul T, Suppakarn N. Role of Maleic Anhydride-Grafted poly(lactic acid) in improving shape memory Properties of Thermoresponsive Poly(ethylene glycol) and poly(lactic acid) blends. *Polymers.* 2022;14(18):3923.
212. Ma X, Zhukov S, von Seggern H, Sessler GM, Ben Dali O, Kupnik M et al. Biodegradable and Bioabsorbable Poly(lactic acid) Ferroelectrets with Prominent Piezoelectric Activity. *Advanced Electronic Materials.* n/a(n/a):2201070. doi: <https://doi.org/10.1002/aelm.202201070>.
213. Shabani Samghabadi M, Karkhaneh A, Katbab AA. Synthesis and characterization of biphasic layered structure composite with simultaneous electroconductive and piezoelectric behavior as a scaffold for bone tissue engineering. *Polymers for Advanced Technologies.* n/a(n/a). doi: <https://doi.org/10.1002/pat.5976>.
214. Dai X, Yao X, Zhang W, Cui H, Ren Y, Deng J, et al. The osteogenic role of Barium Titanate/Poly(lactic acid) Piezoelectric Composite membranes as guiding membranes for bone tissue regeneration. *Int J Nanomedicine.* 2022;17:4339–53. <https://doi.org/10.2147/ijn.S378422>.
215. Phogat K, Ghosh SB, Bandyopadhyay-Ghosh S. Recent advances on injectable nanocomposite hydrogels towards bone tissue rehabilitation. *J Appl Polym Sci.* 2023;140(4):e53362. <https://doi.org/10.1002/app.53362>.
216. Jang W, Mun SJ, Kim S-Y, Bong KW. Controlled growth factor delivery via a degradable poly(lactic acid) hydrogel microcarrier synthesized using degassed micromolding lithography. *Colloids Surf B.* 2023;222:113088. <https://doi.org/10.1016/j.colsurfb.2022.113088>.
217. Rajput M, Nilawar S, Chatterjee K. Embedding Silk Fibroin-Alginate Hydrogel in a 3D-Printed porous poly(lactic acid) bone tissue Scaffold augments stem cell function. *Regenerative Eng Translational Med.* 2023. <https://doi.org/10.1007/s40883-022-00286-7>.
218. Narisepalli S, Salunkhe SA, Chitkara D, Mittal A. Asiaticoside polymeric nanoparticles for effective diabetic wound healing through increased collagen biosynthesis: In-vitro and in-vivo evaluation. *Int J Pharm.* 2023;631:122508. <https://doi.org/10.1016/j.jipharm.2022.122508>.
219. Samatya Yilmaz S, Aytac A. The highly absorbent polyurethane/polylactic acid blend electrospun tissue scaffold for dermal wound dressing. *Polym Bull.* 2023. <https://doi.org/10.1007/s00289-022-04633-0>.
220. Topuz M, Hydroxyapatite. Al₂O₃ reinforced poly- (lactic acid) hybrid coatings on magnesium: characterization, mechanical and in-vitro bioactivity properties. *Surf Interfaces.* 2023;37:102724. <https://doi.org/10.1016/j.surfin.2023.102724>.
221. Cicogna F, Passaglia E, Benedettini M, Oberhauser W, Ishak R, Signori F, et al. Rosmarinic and glycyrrhetic acid-modified layered double hydroxides as functional additives for poly(lactic acid)/Poly(Butylene Succinate) blends. *Molecules.* 2023;28(1):347.
222. Magalhães Brandão R, Roberto Batista L, Elvis de Oliveira J, Bispo Barbosa R, Lee Nelson D, Graças Cardoso M. In vitro and in vivo efficacy of poly(lactic acid) nanofiber packaging containing essential oils from *Ocimum basilicum* L. and *Ocimum gratissimum* L. against *Aspergillus carbonarius* and *Aspergillus niger* in table grapes. *Food Chem.* 2023;400:134087. <https://doi.org/10.1016/j.foodchem.2022.134087>.
223. Hamad K, Kaseem M, Ayyoob M, Joo J, Deri F. Polylactic acid blends: the future of green, light and tough. *Prog Polym Sci.* 2018;85:83–127.
224. Jahangirian H, Lemraski EG, Rafiee-Moghaddam R, Webster TJ. A review of using green chemistry methods for biomaterials in tissue engineering. *Int J Nanomed.* 2018;13:5953.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.